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Bioimpedance-assessed muscle wasting and its relation to nutritional intake during the first week of ICU: a pre-planned secondary analysis of Nutriti Study



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Abstract

Background Muscle mass evaluation in ICU is crucial since its loss is related with long term complications, including physical impairment. However, quantifying muscle wasting with available bedside tools (ultrasound and bioimpedance analysis) must be more primarily understood. Bioimpedance analysis (BIA) provides estimates of muscle mass and phase angle (PA).

The primary aim of this study was to evaluate muscle mass changes with bioimpedance analysis during the first 7 days after ICU admission. Secondary aims searched for correlations between muscular loss and caloric and protein debt.

Methods Patients with an expected ICU-stay ≥ 72 h and the need for artificial nutritional support were evaluated for study inclusion. BIA evaluation of muscle mass and phase angle were performed at ICU admission and after 7 days. Considering the difference between ideal caloric and protein targets, with adequate nutritional macronutrients delivered, we calculated the caloric and protein debt. We analyzed the potential correlation between caloric and protein debt and changes in muscle mass and phase angle.

Results 72 patients from September 1st to October 30th, 2019 and from August 1st to October 30th, 2021 were included in the final statistical analysis. Median age was 68 [59–77] years, mainly men (72%) admitted due to respiratory failure (25%), and requiring invasive mechanical ventilation for 7 [4–10] days. Median ICU stay was 8 [6–12] days. Bioimpedance data at ICU admission and after 7 days showed that MM and PA resulted significantly reduced after 7 days of critically illness, 34.3 kg vs 30.6 kg (p < 0.0001) and 4.90° vs 4.35° (p = 0.0004) respectively. Mean muscle loss was 3.84 ± 6.7 kg, accounting for 8.4% [1–14] MM reduction. Correlation between caloric debt (r = 0.14, p = 0.13) and protein debt (r = 0.18, p = 0.13) with change in MM was absent. Similarly, no correlation was found between caloric debt (r = -0.057, p = 0.631) and protein debt (r = -0.095, p = 0.424) with changes in PA.

Conclusions bioimpedance analysis demonstrated that muscle mass and phase angle were significantly lower after 7 days in ICU. The total amount of calories and proteins does not correlate with changes in muscle mass and phase angle.

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Keywords Critically ill patient, Nutrition, Muscle wasting, ICU acquired weakness, Bioimpedance analysis, Proteins, Caloric debt

Background

Critically ill patients experience acute organ damages and disturbed endocrine and metabolic homeostasis [1]. Activation of the inflammatory response and associated endocrine changes drive hypercatabolism to mobilize substrates for energy production rapidly [2, 3]. In the acute phase of critical illness, muscle proteins form a significant source of energy because muscle proteolysis delivers amino acids that are subsequently used for gluconeogenesis [4]. As a result, skeletal muscle mass rapidly reduces [5]. The severity of muscle wasting is closely associated with illness severity [6, 7]. Muscle wasting is one of the key drivers of ICU acquired weakness (ICU-AW), which not only impairs short-term outcomes but it is also associated with impaired physical abilities and reduced quality of life 5 years after ICU discharge [8].

At the bedside, ultrasound can quantify muscle mass [9]. However, ultrasound only quantifies specific muscle groups, which may not reflect total muscle mass [10, 11]. Moreover, evaluation must be done at the same site for repeated measures to avoid bias, which requires well-trained practitioners [12, 13]. Bioimpedance analysis (BIA) could overcome these limitations, although the technique is not well validated in critically ill patients. There is limited data on the potential of BIA to assess muscle wasting in critical illness.

The primary aim of this study was to assess muscle wasting evaluated with BIA in critically ill patients at ICU admission and after 7 days. A secondary aim was to investigate if the provided energy and protein doses during the first week in ICU were associated with BIAassessed muscle loss.

Material and methods

Setting and design

This is a pre-planned secondary analysis of the NUTRITI STUDY, a single-center prospective observational study that evaluated gastrointestinal dysfunction in critically ill patients [14]. The study was performed at the Department of Intensive Care of the Academic Hospital of Udine, a 1000-bed third-level acute care hospital in the North-East of Italy, and the protocol was approved by Ethics Committee of Friuli-Venezia-Giulia region (CEUR-2019-Os-17). The principles of good clinical practice performed the protocol. Written informed consent was obtained from the patient or patient representative before enrolment in the study. The study protocol was registered on ClinicalTrials.gov (NCT05473546).

All consecutive patients admitted to the general ICU from September 1st to October 30th, 2019 and from August 1st to October 30th, 2021 were evaluated for inclusion. No data were collected between the two study periods due to the COVID-19 pandemic, leading to a lack of available research staff [15].

Inclusion and exclusion criteria

We included patients \geq 18 years old with an expected ICU-stay \geq 72 h and the need for artificial nutritional support, either enteral or parenteral. We excluded those with a pre-planned admission to the ICU as a consequence of elective major surgery, patients who did not require any nutritional support, those with a known gastrointestinal dysfunction (for example, gastrointestinal fistula, chronic diarrhoea or history of malabsorption); patients who tested positive for SARS-CoV2 infection (through PCR nasal swab analysis); those with a severe chronic renal disease (defined as estimated glomerular filtration rate < 30 mL/min) or liver disease (diagnosed cirrhosis and/or patient on a waiting list for liver transplantation); requiring renal replacement therapies (continuous renal replacement therapy [CRRT] or hemodialysis [HD]); patients with body mass index $(BMI) < 16 \text{ kg/m}^2 \text{ or } > 40 \text{ kg/m}^2$; patients with neuromuscular disease; patients without BIA parameters recorded; lastly, patients who refused to undergo BIA evaluation.

Recorded data

The following patient's information was collected: (1) general variables (age, sex, weight, height [supine length measurement: using a flexible measuring tape the length between the vertex of the head and the heel was measured], BMI, admission diagnosis, APACHE II [Acute Physiologic Assessment and Chronic Health Evaluation] score, SOFA [Sequential Organ Failure Assessment] score; (2) clinical variables: mechanical ventilation duration, vasopressor use (continuous infusion for at least 24 h), length of ICU and hospital stay (ICU_{LOS} and HOSP_{LOS} respectively), mortality at 30-90-180 days; (3) nutritional variables (NUTRIC [Nutrition Risk in Critically ill] score at ICU admission, serum albumin-white blood cell-haemoglobincreatinine at ICU admission, cumulative fluid balance after first ICU week (including perspiratio insensibilis calculated as following [intubated patients 0.35 mL/ Kg/h, adding 0.1 mL/Kg/h for every 1 °C above 38 °C; non intubated patients 0.5 mL/Kg/h]; 4) BIA analysis data (resistance [Rz], reactance [Xc], muscle mass [MM], extracellular body water [ECW], phase angle [PA]) at ICU admission and after 7 days.

Data were pseudonymized and collected daily into a dedicated Microsoft Excel[®] sheet (v. 2019, Redmond, WA).

Data were reported according to the STrengthening the Reporting of OBservational studies in Epidemiology (STROBE) statements to improve truthfulness and guarantee clarity.

Bioelectrical impedance analysis

Whole-body impedance data were obtained using a tetrapolar impedance plethysmography (EFG V.3 Akern, Florence, Italy) at ICU admission and at day 7. The bioelectrical parameters of resistance (R) and reactance (Xc) were measured using an electric alternating current flux of 400 μ A and an operating frequency of 50 kHz.

In brief, BIA evaluates some characteristics of tissues in response to an application of alternate current [16]. Electrolyte-rich tissues are highly conductive to electrical current, while anhydrous tissues (like fat) resist the current flow. The opposition to the flow of a current is called resistance (R), while the opposition to a current change due to a capacitor is defined as reactance (X_c). The total opposition to an electrical current by both resistance and reactance is the impedance dimension (I) [17]. Single frequency BIA measures only extracellular water (ECW). Considering the participant's height, age and gender in the regression equations, BIA can estimate lean body mass from impedance values and body water content [18].

Moreover, BIA provides PA, which derives from a phase shift caused by resistance to flow determined by capacitors (i.e., healthy cell membranes) that delay the current's flow [19]. High PA correlates with large quantities of intact cell membranes and body cell mass.

Whole-body impedance measurements were taken according to the standard protocols available [20]. After the patient had remained in a supine position with arms separated from trunk by about 30° and legs separated by about 45° for at least 5 min and after the skin was cleaned to ensure good contact, appropriate electrodes were attached to the right hand and foot, according to the standard protocol of the National Institutes of Health technology assessment conference statements [21]. Enteral nutrition was stopped at least 2 h before measurement.

Nutritional protocol, calories and protein debt calculation

As per our internal protocol and according to ESPEN Guidelines [22], our total energy and protein target is 20 kcal/kg and 1.3 g/kg/day after the first week in the ICU. Unless contraindicated, we aim to reach at least 70% of this target at day 4 after ICU admission. Continuous enteral nutrition is the preferred way to achieve this goal, while parenteral nutrition is considered when nutritional targets are not reached after 7 days.

To enable calculation of an energy and protein deficit, the energy and protein target during the first 7 days was defined as follows: 20% of 20 kcal/kg and of 1.3 g/kg actual body weight on the first day, 30% the second day, 50% at day 3, and at least 70% from day 4 to 7. The difference between the prescribed dose of total calories (Kcal_{TAR}) and proteins (Prot_{TAR}), and the adequate nutritional amount delivered (Kcal_{EFF} and Prot_{EFF}) considering the 7 days together, has been called caloric (Kcal_{DEBT}) and protein (Prot_{DEBT}) deficit respectively according to the following formulas: Kcal_{DEBT}=Kcal_{TAR} – Kcal_{EFF} and Prot_{DEBT}=Prot_{TAR} – Prot_{EFF}.

Primary aim

The primary aim of this study was to measure muscle mass changes evaluated through BIA after 7 days of ICU stay.

Secondary aims

Secondary aims were to investigate if the muscular mass estimated with BIA correlates with -BIA-derived PA, and to investigate the correlation between both $Kcal_{DEBT}$ and $Prot_{DEBT}$ and the change in muscle mass and PA as assessed by BIA.

Statistical analysis

Continuous variables are reported as median (interquartile range [IQR]) or mean (SD) as appropriate, and categorical variables are reported as frequencies and percentages. According to the nature of the variables, differences between groups were assessed via t-test, Chi-squared test, and Fisher's exact test. The normality of the distribution was evaluated by the Shapiro–Wilk test.

Spearman or Pearson's test studied the correlation between variables as appropriate. Data were analysed using R software (The R Foundation), while graphics were made with GraphPad Prism (Version 10.0.39).

A p < 0.05 was statistically significant.



Fig. 1 Study flowchart

Table 1 Patient characteristics at ICU admission

	Overall (N = 72)
General variables	
Age (years)	68 [59–77]
Sex (male)	52 (72%)
Height (m)	1.75 [1.65–1.80]
Weight (Kg)	81 [69–90]
Admission BMI (Kg/m ²)	27 [24–29]
Reason of admission in ICU	
Respiratory failure	18 (25%)
Trauma	13 (18%)
Neurological	14 (19%)
Emergency surgery	10 (14%)
Cardiovascular	7 (10%)
Sepsis*	4 (6%)
Other	6 (8%)
APACHE II score	20 [16–25]
SOFA score	7 [6–8]
NUTRIC score	5 [4–6]
Albumin (mg/dL)	29 [18, 27–36]
WBC 10^3/mm ³	10.26 [8.47–13.36]
Hb (g/dL)	10.3 [9.7–11.72]
Creatinine (mg/dL)	1.22 [0.92–1.65]

BMI, Body Mass Index; ICU, Intensive Care Unit; APACHE II, Acute Physiologic Assessment and Chronic Health Evaluation II; SOFA, Sequential Organ Failure Assessment; NUTRIC, Nutrition Risk in Critically ill; WBC, White Blood Cells

* Sepsis from non-respiratory cause

Results

Patient characteristics

During the study period, 72 patients were included as shown in Fig. 1.

Median age was 68 [59–77] years, mainly men (72%) admitted with respiratory failure (25%) (Table 1). All patients required invasive mechanical ventilation for 7 [4–10] days, and 83% of patients received vasopressor support (Table 2). Median ICU_{LOS} and HOSP_{LOS} was 8 [6–12] and 17 [11–30] days respectively (Table 2). Mortality at 180-days was 28%.

Table 2 Clinical outcomes

Clinical outcomes	
MV duration (days)	7 [4–10]
Vasopressors use (%)	60 (83%)
Cumulative fluid balance at day 7 (mL)	- 1601 [- 3871+876]
MV duration (days)	
ICU _{LOS}	8 [6–12]
HOSP _{LOS}	17 [11–30]
Mortality at 30-days	14 (19%)
Mortality at 90-days	16 (22%)
Mortality at 180-days	20 (28%)

MV, Mechanical Ventilation; ICU, Intensive Care Unit; LOS, Length of stay

Table 3 BIA parameters changes after 7 days of ICU stay

	ICU admission	DAY 7	р
Rz (Ω)	412 [354–504]	409 [351–458]	0.0096
Xc (Ω)	36 [18, 26–45]	32 [18, 23–41]	0.0079
ECW (L)	24.2 [20.2–26.5]	23.9 [20.5–29.1]	0.3463
MM (Kg)	34.3 [27.4–59.6]	30.6 [18, 27–37]	< 0.0001
PA (°)	4.90 [3.65–6.07]	4.35 [3.32–5.47]	0.0004

Rz, resistance; Xc, reactance; ECW, extracellular water; MM, muscle mass; PA, phase angle.

Changes in BIA-assessed muscle mass

BIA data were obtained after 6 [3–11] hours after admission BIA at ICU admission and after 7 days showed that MM and PA significantly reduced after 7 days of critically illness (Table 3). Median MM loss was 2.65 [0.40–5.15] Kg, accounting for 8.4% [1–14] MM reduction after 7 days of ICU stay. A positive correlation was found between MM and PA at admission (r = 0.67, p < 0.0001) as shown in Fig. 2 (left part).

Similarly, changes in MM correlated with changes in PA after 7 days in ICU (r = 0.68, p < 0.0001), see Fig. 2 (right part).

At ICU admission, there was no significant difference in MM nor PA values between patients who subsequently survived versus died during the 180-day follow up period, 32.8 [26.8–44.7] vs 35.2 [28–44.8] Kg (p=0.44) and 4.8° [3.6–6] vs 5.4° [3.7–6] respectively (p=0.58). However, a trend toward higher MM loss was recorded in 180-day non-survivors than in surviving patients, 2.95 kg [1.42–11.53] vs 1.85 kg [0.20–4.67] respectively, p=0.058. Likewise, patients not surviving until 180 days after admission had a greater PA reduction after 7 days of ICU stay than survivors (– 0.70° [– 1.78–0.1] in non-survivors vs -0.15° [– 0.65–0.17] in survivors, p=0.048).



Fig. 2 Correlations between MM and PA at ICU admission and their changes after 7 days of ICU stay. In the left part of the figure, it is shown the correlation between MM and PA at ICU admission (r = 0.67, p < 0.0001): In the right part, MM and PA changes after 7 days from admission demonstrated still to be correlated significantly (r = 0.68, p < 0.0001)

Association of a nutritional deficit with BIA-derived changes in muscle mass and phase angle

The median Kcal_{TAR} to deliver during the first 7 days was 6642 kcal [5740–7380] and the median $Prot_{TAR}$ was 494.9 g [427.7–549.9]. Median Kcal_{EFF} provided during the first 7 days was 3271 kcal [2145–4818] and median $Prot_{EFF}$ was 147.2 g [90.4–256.5], determining a median Kcal_{DEBT} of 3340 kcal [1574–4763] and $Prot_{DEBT}$ of 326 g [214.8–426.7]. There was no correlation between both Kcal_{DEBT} (ρ =0.14, p=0.13) and $Prot_{DEBT}$ (ρ =0.18, p=0.13), and changes in MM over the first week (Fig. 3).

Similarly, there was no correlation between both $\text{Kcal}_{\text{DEBT}}$ (ρ = -0.06, p = 0.63) and $\text{Prot}_{\text{DEBT}}$ (ρ = - 0.09, p = 0.42), and changes in PA (Fig. 4).

Discussion

In this observational study in long-stay critically ill patients, we found that muscle mass assessed by BIA was significantly reduced after 7 days in ICU, with more severe muscle mass loss in patients not surviving the 180-days follow-up period. Muscle mass and changes in muscle mass correlated with BIA-derived phase angle and changes in phase angle respectively. Hence, changes in phase angle could result from changes in muscle mass in critical illness. An early energy and protein deficit did not correlate with changes in muscle mass.

Preventing muscle weakness is important in critical care medicine, as ICU-acquired weakness is associated with physical impairment and reduced quality of life that may persist for a long time after ICU admission [23, 24]. ICU-acquired weakness is provoked by critical



Fig. 3 Correlation between Kcal_{DEBT} and Prot_{DEBT}, and changes in MM after 7 days of ICU stay. No significant correlation was found between variations in MM (Δ MM) and protein debt (Prot_{DEBT}) or caloric debt (Kcal_{DEBT}) during the first 7 days after ICU admission (p=0.13 for both correlations)



Fig. 4 Correlation between Kcal_{DEBT} and Prot_{DEBT}, and changes in PA after 7 days of ICU stay. No significant correlation was found between variations in PA (Δ PA) and protein debt (Prot_{DEBT}) or caloric debt (Kcal_{DEBT}) during the first 7 days after ICU admission (p=0.42 and p=0.63 respectively)

illness polyneuropathy, critical illness myopathy, and loss of muscle mass. In our study we found an 8.4% reduction of MM after 7 days of ICU stay, corresponding to a net 3.84 kg MM loss, which is quite substantial. Although non-survivors had similar muscle mass upon ICU admission, they tended to have a more significant early muscle mass loss than surviving patients, confirming the functional relevance of early muscle mass loss.

Our findings are in line with previous studies that documented substantial early mass loss in critical illness, especially in patients with the high illness severity [25]. Indeed, Puthucheary et al. reported a 12.5% reduction of rectus femoris cross-sectional area after 7 days measured by ultrasound, with increased muscle loss in patients with more severe organ dysfunction [6]. In a meta-analysis, Fazzini et al. reported approximately 2% muscle mass loss per ICU Day in the first week [26]. However, preceding studies primarily evaluated muscular mass changes by ultrasound and computed tomography (CT). Ultrasound only allows evaluation of specific muscle groups, and interrater variability may be high [10]. A CT scan cannot be performed at the bedside and exposes the patient to radiation. Theoretically, these drawbacks of ultrasound and CT could be avoided by BIA assessment of muscle mass [26]. However, the applicability of BIA in critically ill patients has been debated, since increased tissue edema may be a confounding factor [27, 28]. Nevertheless, recent studies suggest the potential usefulness of BIA in critical illness to assess muscle mass. Indeed, 2 studies, altogether including 245 critically ill patients, found a good correlation between BIA-assessed muscle mass and CT findings [29, 30].

The phase angle, a parameter derived from the arctangent value of the ratio of reactance to resistance, is independent of conventional regression equations and it is correlated with muscle quantity and quality [18, 31].

In the present work we found a significant correlation between PA and BIA derived MM at ICU admission (r=0.67, p<0.0001). Further, the correlation between absolute variations of PA and MM during the first week of ICU stay was still present (r=0.68, p<0.0001).

According to the results and the literature, PA significantly decreased after one week in ICU stay with a concomitant reduction in muscle mass [32].

Therefore, monitoring PA variations may be a simple tool to evaluate MM variations in ICU.

The absent correlation between early muscle wasting and a protein and energy deficit confirms anabolic resistance in the acute phase of critical illness. Indeed, despite observational studies associating a higher protein intake with improved outcomes [33], recent large randomized controlled trials have shown potential harm by increased nutritional intake and increased protein doses in the acute phase of critical illness [34–37]. Indeed, the EPaNIC trial showed harm by early parenteral nutrition supplementing insufficient enteral nutrition compared with withholding parenteral nutrition until one week after ICU admission [35]. Likewise, the NUTRIREA-3 trial showed harm by early complete nutrition compared to restrictive feeding through any route [36]. In both RCTs, urea levels increased in the group receiving extra amino acids, suggesting futile catabolism of the supplementary provided amino acids [38]. Likewise, in a secondary analysis of patients included in the EPaNIC study, CT-assessed muscle mass loss was not prevented by early parenteral nutrition [39], and the incidence of weakness even increased [40]. Harm in the EPaNIC trial was subsequently attributed to the increased protein doses, and

to feeding-induced suppression of autophagy, a crucial cellular recovery process [40–42]. Similarly, in the recent post-hoc analysis of the EFFORT trial, increased protein doses did not improve outcome of critically ill patients, and the intervention increased ureagenesis and associated with increased mortality in patients with acute kidney injury and high organ failure scores [43]. The time point when anabolic responsiveness switches into feeding responsiveness remains to be defined and likely varies between patients. BIA may assist in documenting the presence or absence of an anabolic response to artificial nutrition in intervention studies, which requires further study.

Contrary to Thibault et al. [44], we did not observe a significant ICU admission phase angle difference between survivors and non-survivors at 28-day followup. However, this study found that patients not surviving until 180 days after admission had a greater PA reduction than survivors (p=0.048). This may reflect the effect of a higher disease severity, which in turn leads to the patient's death. Moreover, we cannot exclude that the greater PA decrease in non-survivors could result from a maladaptive host response to the acute illness, such as recently shown in critically ill COVID-19 patients [45].

Some limitations should be acknowledged. Firstly, the study is observational, and we did not correct for potential confounders. However, recent randomized controlled trials confirm anabolic resistance in the acute phase of critical illness.

Secondly, we did not assess other markers of muscle mass or function. Future studies should investigate the accuracy and agreement in low muscle mass identification using diverse markers compared to changes in BIAassessed muscle mass [46]. However, the degree of early muscle mass loss was similar to studies assessing muscle wasting by other techniques, and the more significant muscle wasting in non-surviving patients compared to surviving patients supports the functional relevance of our findings.

Third, we did not evaluate the fluid intake from ICU admission to the obtainment of BIA values, which could have altered PA or MM estimation. However, median time from ICU admission to available BIA data was short, and near 50% of patients had an acute illness that did not require large fluidic volume resuscitation.

Lastly, we included a low proportion of septic patients considered at higher risk of muscle derangements than another category of ICU patients [47].

Conclusions

In conclusion, BIA can provide potentially helpful information on muscle wasting in the ICU, and PA may be a valid substitute for MM in the ICU. An Page 7 of 9

early protein and energy deficit did not correlate with increased muscle wasting, supporting the concept of anabolic resistance in the acute phase of critical illness.

Abbreviations

BIA	Bioimpedance analysis
ICU	Intensive care unit
MM	Muscular mass
PA	Phase angle
ICU-AW	ICU acquired weakness
CRRT	Continuous renal replacement therapy
HD	Hemodialysis
BMI	Body mass index
APACHE	Acute Physiologic Assessment and Chronic Health Evaluation
	score
SOFA	Sequential Organ Failure Assessment score
ICU _{LOS}	Length of ICU stay
HOSPLOS	Length of hospital stay
NUTRIC	Nutrition Risk in Critically ill
Rz	Resistance,
Хс	Reactance
ECW	Extracellular body water

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Author contributions

CD, MD and LV conceptualized the study, planned statistical analysis, collected and analyzed data, wrote the manuscript. JG, SDR, MU, DGB, TP, AC and AM helped in analyzing data, writing the manuscript and in critically revising the manuscript and providing important intellectual content. JG shares first authorship with CD. All Authors read and approved the final version of this manuscript. AM and LV share senior authorship.

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Availability of data and materials

The datasets generated and/or analysed during the current study are not publicly available due privacy concerns, but are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

Ethics Committee of Friuli-Venezia-Giulia region approved the study as CEUR-2019-Os-17 also registered on ClinicalTrials.gov (Identifier: NCT05473546, registered on 26th July 2022). Written informed consents were obtained from patients or relatives before enrollment in the study.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- 1. Boonen E, Van den Berghe G. Endocrine responses to critical illness: novel insights and therapeutic implications. J Clin Endocrinol Metab. 2014;99(5):1569–82. https://doi.org/10.1210/jc.2013-4115.
- Preiser JC, Ichai C, Orban JC, Groeneveld AB. Metabolic response to the stress of critical illness. Br J Anaesth. 2014;113(6):945–54. https://doi.org/ 10.1093/bja/aeu187.
- 3. Hamill RW, Woolf PD, McDonald JV, Lee LA, Kelly M. Catecholamines predict outcome in traumatic brain injury. Ann Neurol. 1987;21:438–43.
- Iapichino G, Radrizzani D, Armani S, Noto A, Spanu P, Mistraletti G. Metabolic treatment of critically ill patients: energy balance and substrate disposal. Minerva Anestesiol. 2006;72:533–41.
- Gamrin-Gripenberg L, Sundström-Rehal M, Olsson D, Grip J, Wernerman J, Rooyackers O. An attenuated rate of leg muscle protein depletion and leg free amino acid efflux over time is seen in ICU long-stayers. Crit Care. 2018;22(1):13. https://doi.org/10.1186/s13054-017-1932-6.
- Puthucheary ZA, Rawal J, McPhail M, Connolly B, Ratnayake G, Chan P, et al. Acute skeletal muscle wasting in critical illness. JAMA. 2013;310(15):1591–600. https://doi.org/10.1001/jama.2013.278481.
- Wandrag L, Brett SJ, Frost GS, Bountziouka V, Hickson M. Exploration of muscle loss and metabolic state during prolonged critical illness: Implications for intervention? PLoS ONE. 2019;14(11): e0224565. https://doi.org/ 10.1371/journal.pone.0224565.
- Vanhorebeek I, Latronico N, Van den Berghe G. ICU-acquired weakness. Intensive Care Med. 2020;46(4):637–53. https://doi.org/10.1007/ s00134-020-05944-4.
- De Rosa S, Umbrello M, Pelosi P, Battaglini D. Update on lean body mass diagnostic assessment in critical illness. Diagnostics. 2023;13(5):888. https://doi.org/10.3390/diagnostics13050888.
- Paris MT, Lafleur B, Dubin JA, Mourtzakis M. Development of a bedside viable ultrasound protocol to quantify appendicular lean tissue mass. J Cachexia Sarcopenia Muscle. 2017;8(5):713–26. https://doi.org/10.1002/ jcsm.12213.
- Formenti P, Umbrello M, Coppola S, Froio S, Chiumello D. Clinical review: peripheral muscular ultrasound in the ICU. Ann Intensive Care. 2019;9(1):57. https://doi.org/10.1186/s13613-019-0531-x.
- Mourtzakis M, Parry S, Connolly B, Puthucheary Z. Skeletal muscle ultrasound in critical care: a tool in need of translation. Ann Am Thorac Soc. 2017;14(10):1495–503. https://doi.org/10.1513/AnnalsATS.201612-967PS.
- Curz-Jentoft AJ, Bahat G, Bauer J, Boirie Y, Bruyere O, Cederholm T, et al. Sarcopenia: revised European consensus on definition and diagnosis. Age Ageing. 2019;48:16–31.
- 14. Danielis M, Mattiussi E, Piani T, Iacobucci A, Tullio A, Molfino A, Nutriti Study Group, et al. Diarrhoea and constipation during artificial nutrition

in intensive care unit: a prospective observational study. Clin Nutr ESPEN. 2023;57:375–80. https://doi.org/10.1016/j.clnesp.2023.07.007.

- Deana C, Rovida S, Orso D, Bove T, Bassi F, De Monte A, et al. Learning from the Italian experience during COVID-19 pandemic waves: be prepared and mind some crucial aspects. Acta Biomed. 2021;92(2): e2021097. https://doi.org/10.23750/abm.v92i2.11159.
- Moonen HPFX, Van Zanten ARH. Bioelectric impedance analysis for body composition measurement and other potential clinical applications in critical illness. Curr Opin Crit Care. 2021;27(4):344–53. https://doi.org/10. 1097/MCC.00000000000840.
- Mulasi U, Kuchnia AJ, Cole AJ, Earthman CP. Bioimpedance at the bedside: current applications, limitations, and opportunities. Nutr Clin Pract. 2015;30(2):180–93. https://doi.org/10.1177/0884533614568155.
- Lukaski HC. Evolution of bioimpedance: a circuitous journey from estimation of physiological function to assessment of body composition and a return to clinical research. Eur J Clin Nutr. 2013;67(suppl 1):S2–9.
- Myatchin I, Abraham P, Malbrain MLNG. Bio-electrical impedance analysis in critically ill patients: are we ready for prime time? J Clin Monit Comput. 2020;34(3):401–10. https://doi.org/10.1007/s10877-019-00439-0.
- Lukaski HC, Bolonchuk WW, Hall CB, Siders WA. Validation of tetrapolar bioelectrical impedance method to assess human body composition. J Appl Physiol. 1986;60(4):1327–32. https://doi.org/10.1152/jappl.1986.60.4. 1327.
- Bioelectrical Impedance Analysis in Body Composition Measurement. Proceedings of a National Institutes of Health Technology Assessment Conference. Bethesda, Maryland, December 12-14, 1994. Am J Clin Nutr. 1996;64(3 Suppl):387S-532S. https://doi.org/10.1093/ajcn/64.3.387S.
- Singer P, Blaser AR, Berger MM, Alhazzani W, Calder PC, Casaer MP, et al. ESPEN guideline on clinical nutrition in the intensive care unit. Clin Nutr. 2019;38(1):48–79. https://doi.org/10.1016/j.clnu.2018.08.037.
- Deana C, Vetrugno L, Cortegiani A, Mongodi S, Salve G, Mangiagalli M, et al. On behalf of the Italian Odissea group quality of life in COVIDrelated ards patients one year after intensive care discharge (Odissea Study): a Multicenter Observational Study. J Clin Med. 2023;12(3):1058. https://doi.org/10.3390/jcm12031058.
- Herridge MS, Tansey CM, Matté A, Tomlinson G, Diaz-Granados N, Cooper A, et al. Canadian Critical Care Trials Group. Functional disability 5 years after acute respiratory distress syndrome. N Engl J Med. 2011;364(14):1293–304. https://doi.org/10.1056/NEJMoa1011802.
- 25. Lopez-Ruiz A, Kashani K. Assessment of muscle mass in critically ill patients: role of the sarcopenia index and images studies. Curr Opin Clin Nutr Metab Care. 2020;23(5):302–11. https://doi.org/10.1097/MCO.00000 00000000673.
- Fazzini B, Märkl T, Costas C, Blobner M, Schaller SJ, Prowle J, et al. The rate and assessment of muscle wasting during critical illness: a systematic review and meta-analysis. Crit Care. 2023;27(1):2. https://doi.org/10.1186/ s13054-022-04253-0.
- Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Manuel Gómez J, et al. ESPEN. Bioelectrical impedance analysis-part II: utilization in clinical practice. Clin Nutr. 2004;23(6):1430–53. https://doi.org/10.1016/j.clnu. 2004.09.012.
- Forni LG, Hasslacher J, Joannidis M. Bioelectrical impedance vector analysis in the critically ill: cool tool or just another 'toy'? Crit Care. 2015;11(19):387. https://doi.org/10.1186/s13054-015-1110-7.
- Kim D, Sun JS, Lee YH, Lee JH, Hong J, Lee JM. Comparative assessment of skeletal muscle mass using computerized tomography and bioelectrical impedance analysis in critically ill patients. Clin Nutr. 2019;38(6):2747–55. https://doi.org/10.1016/j.clnu.2018.12.002.
- Looijaard WGPM, Stapel SN, Dekker IM, Rusticus H, Remmelzwaal S, Girbes ARJ, et al. Identifying critically ill patients with low muscle mass: agreement between bioelectrical impedance analysis and computed tomography. Clin Nutr. 2020;39(6):1809–17. https://doi.org/10.1016/j.clnu. 2019.07.020.
- Norman K, Stobäus N, Pirlich M, Bosy-Westphal A. Bioelectrical phase angle and impedance vector analysis–clinical relevance and applicability of impedance parameters. Clin Nutr. 2012;31(6):854–61. https://doi.org/ 10.1016/j.clnu.2012.05.008.
- Formenti P, Coppola S, Umbrello M, Froio S, Caccioppola A, De Giorgis V, et al. Time course of the bioelectrical impedance vector analysis and muscular ultrasound in critically ill patients. J Crit Care. 2022;68:89–95. https://doi.org/10.1016/j.jcrc.2021.11.014.

- 33. Weijs PJ, Looijaard WG, Beishuizen A, Girbes AR, Oudemans-van Straaten HM. Early high protein intake is associated with low mortality and energy overfeeding with high mortality in non-septic mechanically ventilated critically ill patients. Crit Care. 2014;18(6):701. https://doi.org/10.1186/ s13054-014-0701-z.
- Heyland DK, Patel J, Compher C, Rice TW, Bear DE, Lee ZY, et al. EFFORT protein trial team the effect of higher protein dosing in critically ill patients with high nutritional risk (EFFORT Protein): an international, multicentre, pragmatic, registry-based randomised trial. Lancet. 2023;401(10376):568–76. https://doi.org/10.1016/S0140-6736(22) 02469-2
- Casaer MP, Mesotten D, Hermans G, Wouters PJ, Schetz M, Meyfroidt G, et al. Early versus late parenteral nutrition in critically ill adults. N Engl J Med. 2011;365(6):506–17. https://doi.org/10.1056/NEJMoa1102662.
- Reignier J, Plantefeve G, Mira JP, Argaud L, Asfar P, Aissaoui N, et al. NUTRIREA-3 Trial Investigators; Clinical Research in Intensive Care; Sepsis (CRICS-TRIGGERSEP)Group. Low versus standard calorie and protein feeding in ventilated adults with shock: a randomised, controlled, multicentre, open-label, parallel-group trial (NUTRIREA-3). Lancet Respir Med. 2023. https://doi.org/10.1016/S2213-2600(23)00092-9.
- Fivez T, Kerklaan D, Mesotten D, Verbruggen S, Wouters PJ, Vanhorebeek I, et al. Early versus Late Parenteral Nutrition in Critically III Children. N Engl J Med. 2016;374(12):1111–22. https://doi.org/10.1056/NEJMoa1514762. (Epub 2016 Mar 15).
- Jan G, Ilse V, Thiessen Steven E, Van den Berghe G. Amino acid supplements in critically ill patients. Pharmacol Res. 2018. https://doi.org/10. 1016/j.phrs.2017.12.007.
- Casaer MP, Langouche L, Coudyzer W, Vanbeckevoort D, De Dobbelaer B, Güiza FG, et al. Impact of early parenteral nutrition on muscle and adipose tissue compartments during critical illness. Crit Care Med. 2013;41(10):2298–309. https://doi.org/10.1097/CCM.0b013e31828cef02.
- Hermans G, Casaer MP, Clerckx B, Güiza F, Vanhullebusch T, Derde S, et al. Effect of tolerating macronutrient deficit on the development of intensive-care unit acquired weakness: a subanalysis of the EPaNIC trial. Lancet Respir Med. 2013;1(8):621–9. https://doi.org/10.1016/S2213-2600(13)70183-8.
- Casaer MP, Wilmer A, Hermans G, Wouters PJ, Mesotten D, Van den Berghe G. Role of disease and macronutrient dose in the randomized controlled EPaNIC trial: a post hoc analysis. Am J Respir Crit Care Med. 2013;187(3):247–55. https://doi.org/10.1164/rccm.201206-0999OC.
- Gunst J. Recovery from critical illness-induced organ failure: the role of autophagy. Crit Care. 2017;21(1):209. https://doi.org/10.1186/ s13054-017-1786-y.
- Stoppe C, Patel JJ, Zarbock A, Lee ZY, Rice TW, Mafrici B, et al. The impact of higher protein dosing on outcomes in critically ill patients with acute kidney injury: a post hoc analysis of the EFFORT protein trial. Crit Care. 2023;27(1):399. https://doi.org/10.1186/s13054-023-04663-8.
- 44. Thibault R, Makhlouf AM, Mulliez A, Cristina Gonzalez M, Kekstas G, Kozjek NR, et al. Phase Angle Project Investigators. Fat-free mass at admission predicts 28-day mortality in intensive care unit patients: the international prospective observational study Phase Angle Project. Intensive Care Med. 2016;42(9):1445–53. https://doi.org/10.1007/s00134-016-4468-3.
- Dorward DA, Russell CD, Um IH, Elshani M, Armstrong SD, Penrice-Randal R, et al. Tissue-specific immunopathology in fatal COVID-19. Am J Respir Crit Care Med. 2021;203(2):192–201. https://doi.org/10.1164/rccm. 202008-3265OC.
- Umbrello M, Guglielmetti L, Formenti P, Antonucci E, Cereghini S, Filardo C, et al. Qualitative and quantitative muscle ultrasound changes in patients with COVID-19-related ARDS. Nutrition. 2021;91:111449. https:// doi.org/10.1016/j.nut.2021.111449.
- Callahan LA, Supinski GS. Sepsis-induced myopathy. Crit Care Med. 2009;37(10 Suppl):S354–67. https://doi.org/10.1097/CCM.0b013e3181 b6e439.

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