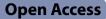
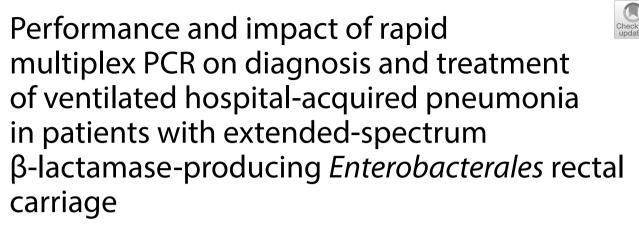
## RESEARCH

Annals of Intensive Care





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## Abstract

**Background** Antimicrobial stewardship (AMS) for ventilator-associated pneumonia (VAP) or ventilated hospitalacquired pneumonia (vHAP) in extended-spectrum  $\beta$ -lactamase-producing *Enterobacterales* (ESBL-E) carriers is challenging. BioFire<sup>®</sup> FilmArray<sup>®</sup> Pneumonia plus Panel (mPCR) can detect bacteria and antibiotic resistance genes, including *bla*<sub>CTX-M</sub>, the most common ESBL-encoding gene.

**Methods** This monocentric, prospective study was conducted on a group of ESBL-E carriers from March 2020 to August 2022. The primary objective was to evaluate the concordance between the results of mPCR and conventional culture performed on respiratory samples of ESBL-E carriers to investigate suspected VAP/vHAP. The secondary objective was to appraise the impact of performing or not mPCR on initial antibiotic therapy adequacy in ESBL-E carriers with confirmed VAP/vHAP.

**Results** Over the study period, 294 patients with ESBL-E carriage were admitted to the ICU, of who 168 (57%) were mechanically ventilated. (i) Diagnostic performance of mPCR was evaluated in suspected 41 episodes of VAP/vHAP:  $bla_{CTX-M}$  gene was detected in 15/41 (37%) episodes, where 9/15 (60%) were confirmed ESBL-E-induced pneumonia. The culture and  $bla_{CTX-M}$  were concordant in 35/41 (85%) episodes, and in all episodes where  $bla_{CTX-M}$  was negative (n = 26), the culture never detected ESBL-E. (ii) The impact of mPCR on initial antibiotic therapy adequacy was assessed in 95 episodes of confirmed VAP/vHAP (22 episodes were tested with mPCR and 73 without); 47 (49%) episodes were ESBL-E-induced, and 24 (25%) were carbapenem-resistant bacteria-induced. The use of mPCR was significantly associated with higher prescription of adequate empirical antibiotic therapy in the multivariable logistic regression

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(adjusted odds ratio (aOR) (95% Cl) of 7.5 (2.1–35.9), p = 0.004), propensity-weighting (aOR of 5.9 (1.6–22.1), p = 0.008), and matching-cohort models (aOR of 5.8 (1.5–22.1), p = 0.01).

**Conclusion** mPCR *bla*<sub>CTX-M</sub> showed an excellent diagnostic value to rule out the diagnosis of ESBL-E related pneumonia in ESBL-E carriers with suspected VAP/vHAP. In addition, in patients with confirmed VAP/vHAP, a mPCR-based antibiotic therapy was associated with an increased prescription of adequate empirical antibiotic therapy. Performing mPCR on respiratory samples seems to be a promising tool in ESBL-E carriers with suspected vHAP/VAP. However, if mPCR is used in very low pre-test clinical probability of pneumonia, due to the high sensitivity and the rate of overdiagnosed pneumonia, the risk of overconsumption of carbapenem may prevail. Further studies are warranted.

**Keywords** Ventilator-associated pneumonia, Multiplex PCR, Antimicrobial stewardship, ESBL, Nosocomial pneumonia, Carbapenem, Intensive care unit

## Introduction

The most common indication for antibiotic treatment in the intensive care unit (ICU) that accounts for half of its prescriptions is suspected lower respiratory tract infection [1]. The need for early use of adequate antibiotic regimen in the ICU should be weighed against the risk of promoting multidrug-resistant (MDR) bacteria via unnecessary broad spectrum antibiotic therapy [2]. Antimicrobial stewardship (AMS) is even more challenging in patients whose digestive tracts are colonised with extended-spectrum β-lactamase-producing *Enterobacte*rales (ESBL-E), a known risk factor for infections [2]. The French guidelines recommend the use of carbapenems for suspected ventilator-associated pneumonia (VAP) in ESBL-E colonised patients who are immunosuppressed or presenting signs of severity [3]. However, ESBL-E related VAP accounted for only 7% of infection-related ventilator-associated complications in ESBL-E carriers, making carbapenems prescription often unnecessary [4]. The lack of reliable predictor of ESBL-E-related pneumonia in ESBL-E carriers and the relatively high prevalence of pneumonia caused by carbapenem-resistant bacteria (CRB) in ESBL-E carriers are strong arguments to look for novel diagnostic approaches [4].

BioFire<sup>®</sup> FilmArray<sup>®</sup> Pneumonia plus Panel (bioMérieux, France) is a rapid multiplex PCR (mPCR) test that can detect in 1.5 h, when performed on respiratory samples, 18 bacteria, nine viruses, and seven antibiotic resistance genes, including *bla*<sub>CTX-M</sub>, the most widely represented ESBLs in *Enterobacterales* isolated in the USA and Europe today [5]. Despite its good diagnostic value [6–10], mPCR showed conflicting results on AMS [11, 12] and has never been tested in ESBL-E carriers, a specific population with high risk of ESBL-E-related infections.

The primary objective of this study was to evaluate the concordance between the results of mPCR and conventional culture applied on respiratory samples of ESBL-E carriers with suspected VAP/vHAP. The secondary objective was to appraise the impact of performing or not

mPCR on initial antibiotic therapy adequacy in ESBL-E carriers with confirmed vHAP/VAP.

## Methods

#### Setting and patients

This monocentric observational prospective study, was conducted from March 2020 to August 2022 in a medical ICU of a university hospital. We included all ESBL-E carriers receiving invasive mechanical ventilation for more than 2 days and those requiring invasive mechanical ventilation for hospital-acquired pneumonia (i.e., vHAP). Intestinal carriage of ESBL-E was screened by rectal swabbing at ICU admission and weekly afterwards. The following data were collected: age, sex, comorbidities, Simplifed Acute Physiology Score (SAPS II), main reason for admission, antibiotic class received during ICU stay, clinical and biological features at time of sampling, and empirical antibiotic class initiated after sampling, after mPCR results, after quantitative culture results and after antibiotic susceptibility testing (AST) results.

Pneumonia was clinically suspected upon discovering new or persistent pulmonary infiltrates on chest X-ray associated with two of the following: purulent respiratory secretions, fever or hypothermia (body temperature greater > 38 or < 36 °C, respectively), leukocytosis or leukopenia (white blood cells count  $\geq 12 \times 10^9$  or  $\leq 4 \times 10^9/L$ , respectively) [4, 13]. Confirmed pneumonia was defined by quantitative culture from a protected telescopic catheter samples ( $\geq 10^3$  CFU/mL), bronchoalveolar lavage fluid ( $\geq 10^4$  CFU/mL), or endotracheal aspirate  $(\geq 10^5 \text{ CFU/mL})$ . These thresholds were not applied to mPCR results. Noteworthy, the BioFire Pneumonia test was not initially validated on protected telescopic catheter, but recent studies have evidenced its good diagnostic value on such samples [8, 9, 14, 15]. VAP was defined as pneumonia developing after  $\geq$  48 h of endotracheal intubation, whereas vHAP was defined as pneumonia occurring within the 24 h preceding intubation in patients hospitalised for at least 48 h [16].

#### **Microbiological analysis**

Conventional microbiological analyses were conducted in compliance with EUCAST recommendations and included quantitative culture, bacterial identifications using Matrix-Assisted Laser Desorption/Ionisation-Time-Of-Flight mass spectrometer (Microflex LT, Bruker Daltonics, Bremen, Germany), and AST performed using disk diffusion method on Mueller-Hinton media (Bio-Rad, Marnes-la-Coquette, France) on colonies isolated after the primary culture. In Enterobacterales, ESBL were phenotypically detected on AST if a difference of more than 5 mm was observed between the discs "Cefepime" and "Cefepime+clavulanate" and/or using a double-disk synergy test [17]. A carbapenemase was phenotypically suspected on AST when the ertapenem diameter was below the susceptibility breakpoint and confirmed by qualitative lateral flow immunoassay (NG-Test® CARBA-5, NG-Biotech, Guipry, France). FilmArray<sup>®</sup> Pneumonia plus panel was implemented according to the manufacturer's instructions using 200 µL of the mucolytic SLdiluted solution (Copan) as a sample for the pouch-based mPCR with FilmArray Torch instrument [18]. Intensivists obtained the results of mPCR 24/7 and within two hours from receiving the sample at the laboratory. mPCR was performed whatever direct smear examination results. For endotracheal aspirates, mPCR was performed only if there was polymorphonuclear cells without squamous epithelial cells.

# Diagnostic performance of mPCR *bla*<sub>CTX-M</sub> in ESBL-E carriers with suspected of VAP/vHAP

The primary objective of the study was to evaluate prospectively the concordance between the results of ESBL-E quantitative culture and mPCR/bla<sub>CTX-M</sub> tests, performed on respiratory samples of ESBL-E carriers suspected to have vHAP/VAP. For each micro-organism identification, a result was considered true positive (TP) or true negative (TN) if the results of mPCR and conventional techniques were concordant in that purpose. The conventional cultures were considered as the reference method, i.e., a microorganism identified only by the mPCR and not by the conventional techniques was considered as a false positive (FP), and conversely a target found by the conventional methods and not by the mPCR, was considered a false negative (FN). Agreement between the two methods was assessed by calculating the positive percentage agreement (PPA), and the negative percentage agreement (NPA) rather than sensitivity and specificity as it was difficult to count on standard culture methods as the gold standard [6, 19–21]. PPA was calculated as (TP/(TP + FN)) and NPA as (TN/(TN + FP)). The positive predictive value and the negative predictive value were calculated as  $100^{(TP/(TP+FP))}$  and  $100^{(TN/(TN+FN))}$ , respectively. Accuracy was calculated as (TP+TN)/(TP+TN+FN+FP).

#### Impact of mPCR results on initial antibiotic therapy adequacy in ESBL-E carriers with confirmed VAP/vHAP

The secondary objective was to assess retrospectively the impact of using mPCR (mPCR group) on initial antibiotic therapy in ESBL-E carriers with confirmed vHAP/ VAP versus conventional diagnostic strategy without mPCR (conventional group). Briefly, mPCR was performed at the physician's discretion and empirical antibiotic therapy was based on a restrictive antibiotic policy [22] and guidelines [3, 23]. No repetition of mPCR was performed for the same episode. Our ICU protocol for empirical antibiotic therapy is provided in the supplementary methods (Supplementary 1). The clinical impact of mPCR was assessed by the rate of empirical therapies retained as adequate and optimal. Empirical antibiotic therapy referred to the antibiotics prescribed before obtaining quantitative culture results (i.e., after sampling, gram coloration and obtaining mPCR results in the mPCR group, and after sampling and gram coloration in the conventional group). The empirical antibiotic therapy was considered adequate if at least one agent was active against all causative pathogens identified by the conventional microbiological culture, based on AST findings. On the other hand, the therapy was considered optimal if the active agent had the narrowest possible spectrum (Supplementary 1 [24]). The time required to designate optimal antibiotic therapy was defined as the interval between drawing the respiratory sample on which the diagnosis of pneumonia was made, and the initiation of optimal antibiotic therapy, expressed in hours.

#### Statistical analysis

Categorical variables, expressed as number (%), were compared using Chi-square or Fisher's exact tests, whereas continuous variables, expressed as median [25–75th percentile interquartile range (IQR)], were compared using Student's t-test or Wilcoxon's rank test, as appropriate. To identify characteristics of episodes associated with adequate empiric antibiotics therapy in patients with confirmed VAP/vHAP, we used multivariable logistic regression. Non-redundant variables selected in bivariate analysis (p<0.10) and considered clinically relevant were entered into the logistic regression model. To rule out indication biases related to the use of mPCR, multivariable analyses were conducted using overlap propensity-score weighting and propensity-score matching methods. Confounders included in the propensity score were the three following patients' characteristics recorded at time of sampling: circulatory failure defined as cardiovascular SOFA score of  $\geq$  3, ratio of partial pressure of arterial oxygen to fraction of inspired oxygen (PaO2/FiO2) of <150 mmHg, and the use of carbapenem within the 72 h prior to sampling (a known protective factor against ESBL-E pneumonia) [4]. Standardised mean differences were examined to assess balance between groups before and after weighting and matching (eFigure 1). R scripts are provided as supplementary material (Supplementary 1). Statistical significance was defined as P<0.05. Analyses were computed with IBM SPSS Statistics v22.0 software (IBM Corp, Armonk, NY) and RStudio software, version 4.2.0 (https://www.R-project. org/). The methods and results of this study are presented according to the STROBE guidelines [25].

#### **Ethical considerations**

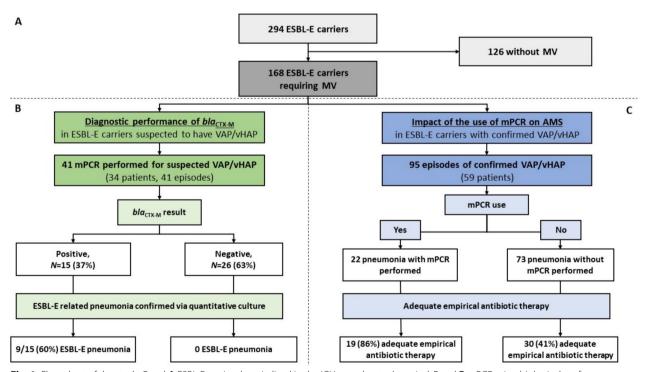
This observational study was approved by the Institutional Review Board of Henri Mondor university hospital and its database registered by the "*Commission Nationale de l'Informatique et des Libertés*" (n°2,232,944). Patients were informed of their inclusion in the study and written informed consent was waived as per French law.

#### Results

Over the study period, 2827 patients required ICU admission. Of them, 1497 patients had at least one ESBL-E screening by rectal swab, and 294 (10.4%) had a positive rectal swab for ESBL-E. 168 ESBL-E rectal carriers required mechanical ventilation (Fig. 1A). The primary endpoint (diagnostic performance of mPCR) was evaluated in 41 suspected episodes of VAP/vHAP (Fig. 1B). The secondary endpoint (impact of performing or not mPCR on initial antibiotic therapy adequacy) was assessed in 95 episodes of quantitative culture-confirmed VAP/vHAP (Fig. 1C).

## Diagnostic performance of mPCR in ESBL-E carriers suspected to have VAP/vHAP

Overall, mPCR was performed on the respiratory samples of 34 of the 168 ESBL-E carriers requiring mechanical ventilation (20%), which represents 41 episodes of suspected VAP/vHAP. The characteristics of the patients (N=34) at ICU admission and those of the episodes (N=41) of suspected pneumonia are respectively reported in eTable 1 and Table 1. mPCR was performed on protected telescopic catheter samples (n=28/41, 68%), bronchoalveolar lavage fluids (n=9/41, 22%), and endotracheal aspirates (n=4/41, 10%). *Bla*<sub>CTX-M</sub> gene



**Fig. 1** Flow chart of the study. Panel **A** ESBL-E carriers hospitalized in the ICU over the study period. Panel **B** mPCR microbiological performance in ESBL-E carriers with suspected vHAP/VAP. Panel **C** Impact of the use of mPCR on the decision making to initiate antibiotic therapy in ESBL-E carriers with confirmed vHAP/VAP. *CTX-M* Cefotaximase-Munich, *ESBL-E* extended-spectrum β-lactamase-producing *Enterobacterales, mPCR* multiplex polymerase chain reaction, *VAP* ventilator associated pneumonia, *vHAP* ventilated hospital-acquired pneumonia

 Table 1
 Characteristics of the 41 episodes of suspected vHAP/

 VAP at the time of BioFire<sup>®</sup> FilmArray<sup>®</sup> Pneumonia Panel plus (mPCR)

| Variable   | All episodes, n=41 |
|--|--------------------|
| Days after ICU admission                             | 14 [7–21]          |
| Days after mechanical ventilation                    | 11 [4–18]          |
| ESBL-Enterobacterales colonisation                   |                    |
| Escherichia coli alone                               | 21 (51)            |
| Klebsiella pneumoniae and/or Enterobacter<br>cloacae | 20 (49)            |
| Days after first positive ESBL-E carriage test       | 6 [3–13]           |
| Previous VAP   | 17 (41)            |
| Number of previous VAP                               | 1 [1, 2]           |
| Type of suspected episode                            |                    |
| vHAP   | 6 (15)             |
| VAP  | 35 (85)            |
| Patient clinical characteristics                     |                    |
| Extracorporeal membrane oxygenation                  | 5 (12)             |
| SOFA score   | 8 [5–11]           |
| PaO <sub>2</sub> /FiO <sub>2</sub> , mmHg            | 150 [79–205]       |
| Circulatory failure <sup>1</sup>                     | 26 (63)            |
| Antibiotics received within 72 h prior to mPCR       | 29 (71)            |
| Carbapenem received within 72 h prior to mPCR        | 5 (12)             |

ESBL-E extended-spectrum β-lactamase-producing Enterobacterales, ICU Intensive Care Unit, mPCR multiplex polymerase chain reaction, PaO<sub>2</sub>/FiO<sub>2</sub> ratio of the partial pressure of arterial oxygen to the fraction of inspired oxygen, SOFA sequential organ failure assessment, VAP ventilator associated pneumonia, vHAP ventilated hospital-acquired pneumonia

Continuous variables are expressed as median [interquartile range]; categorical variables are expressed as n (%)

<sup>1</sup> Circulatory failure is defined as cardiovascular SOFA score of  $\geq$  3

was detected in 15/41 (37%) episodes (Fig. 1). Twenty four episodes (59%) had a positive mPCR, of which 20 (83%) with a definite diagnosis of pneumonia. Among the 17 episodes (41%) with a negative mPCR, 2 (12%) had a definite diagnosis of pneumonia. Assessment of mPCR performance in detecting bacterial and resistance genes in comparison with culture is shown in eTable 2. Overall, the results of quantitative culture and  $bla_{\text{CTX-M}}$  were concordant in 35/41 episodes (85%). Noteworthy, when bla<sub>CTX-M</sub> was negative, culture never found an ESBL-E, suggesting that no pneumonia was due to TEM- or SHV-producing isolates. The six episodes with discordance between genotype (mPCR) and phenotype (culture) are detailed in Supplementary 2. In most episodes (n=31/41, 76%), the patients were put on empirical antibiotic therapy immediately after drawing the respiratory sample and before having the mPCR results. All of the 24 episodes with positive mPCR were treated with empirical antibiotic therapy after obtaining the mPCR result, and 19 (79%) of them received carbapenems. Of the remaining 17 episodes where mPCR failed to detect bacteria, 11 (65%) received empirical antibiotic therapy, of which 2 (12%) received carbapenems. The latter antibiotics were systematically used whenever the  $bla_{CTX-M}$  results were positive (n=15/15, 100%), and spared otherwise in most episodes (n=20/26, 77%, p<0.001). An exploratory analysis conducted during the same period on 228 mPCR performed on mechanically ventilated patients with a negative rectal swab for ESBL-E carriage found that mPCR was positive for  $bla_{CTX-M}$  in two patients (one false positive and one true positive).

#### Impact of mPCR use on initial antibiotic therapy adequacy in ESBL-E carriers with confirmed vHAP/VAP

Over the entire study period, 59 ESBL-E carriers developed 95 confirmed vHAP/VAP episodes, of which 22 episodes were tested using mPCR (Fig. 1C). Retrospectively, the identified reasons for not performing mPCR were as follows: the pre-test probability of pneumonia was assessed as low or very low by the clinician in 38 (52%) episodes, a poor quality of sample without leukocytes was present in 11 episodes (15%), 6 episodes (8%) were included at the start of the implementation period of mPCR, and for the remaining 18 episodes (25%), the reason was not reported in the medical record. Patients' characteristics and organ failure during ICU stay are respectively reported in eTable 3 and Table 2. The mPCR group patients had more circulatory failure, higher SOFA score, and were not put on carbapenem within the 72 h prior to sampling, as compared with their counterparts (Table 2). Forty-seven (49%) vHAP/VAP were related to an ESBL-E, with no difference according to using mPCR [38/73 (52%) vs. 9/22 (41%), p=0.4] (eTable 4) and 24 (25%) episodes were CRB-induced. The use of empirical antibiotic therapy was not statistically different between mPCR group and conventional group after sampling (Table 3). The empirical antibiotic therapy was more frequently adequate and optimal for vHAP/VAP for patients in the mPCR group, as compared to their counterparts: 19/22 (86%) vs. 30/73 (41%), p<0.001, and 15/22 (68%) vs. 20/73 (27%), p=0.001, respectively. This effect was more pronounced in ESBL-E related pneumonia. Sensitivity analyses excluding vHAP, episodes for which carbapenems were administered within the 72 h prior to sampling or including the first episode of pneumonia yielded similar results (Table 3). Figure 2 depicts antibiotic therapy stewardship after sampling and mPCR results. The use of mPCR test, having circulatory failure, and low PaO2/FiO2 ratio were significantly associated with prescription of adequate empirical antibiotic therapy, as shown in the univariate analysis (eTable 5). Alike, mPCR testing was significantly associated with adequate empirical antibiotic therapy in the multivariable logistic regression (adjusted odds ratio (aOR) (95% CI)

| Variable   | Conventional group, n = 73 | mPCR group, n = 22 | р     |
|--|----------------------------|--------------------|-------|
| Days after admission to the ICU                    | 25 [10–60]                 | 18 [12–38]         | 0.3   |
| Days after mechanical ventilation                  | 24 [9–59]                  | 13 [9–33]          | 0.1   |
| ESBL Enterobacterales colonisation                 |                            |                    |       |
| Escherichia. Coli alone                            | 32 (44)                    | 11 (50)            | 0.6   |
| Klebsiella Pneumoniae and/or Enterobacter Cloacae  | 36 (49)                    | 11 (50)            | 1     |
| Others <sup>1</sup>                                | 5 (7)                      | 0                  | 0.6   |
| Days after first positive ESBL-E carriage test     | 11 [4–29]                  | 9 [3–17]           | 0.4   |
| Previous VAP                                       | 44 (60)                    | 12 (54)            | 0.6   |
| Number of previous VAP                             | 2 [1-4]                    | 2 [1, 2]           |       |
| Antibiotics received within 72 h prior to sampling | 45 (62)                    | 12 (54)            | 0.6   |
| Carbapenem received within 72 h prior to sampling  | 17 (23)                    | 0                  | 0.01  |
| Type of suspected episode                          |                            |                    | 0.05  |
| VHAP   | 0                          | 2 (9)              |       |
| VAP  | 73 (100)                   | 20 (91)            |       |
| Patient characteristics                            |                            |                    |       |
| Extracorporeal membrane oxygenation                | 23 (31)                    | 5 (23)             | 0.4   |
| SOFA score   | 6 [4–9]                    | 10 [7–11]          | 0.007 |
| PaO <sub>2</sub> /FiO <sub>2</sub> , mmHg          | 151 [83–240]               | 91 [62–185]        | 0.1   |
| PaO2/FiO2 < 150 mmHg                               | 35 (48)                    | 14 (64)            | 0.2   |
| Circulatory failure <sup>2</sup>                   | 30 (41)                    | 15 (68)            | 0.03  |
| Antibiotic therapy on the day of sampling          | 32 (44)                    | 7 (32)             | 0.3   |
| Non-carbapenem β-lactam                            | 23 (31)                    | 7 (32)             | 1     |
| Carbapenem   | 9 (12)                     | 0                  | 0.1   |
| Pneumonia characteristics                          |                            |                    |       |
| ESBL-E related pneumonia                           | 38 (52)                    | 9 (41)             | 0.4   |
| Carbapenem-resistant pneumonia                     | 19 (26)                    | 5 (23)             | 0.8   |

## Table 2 Characteristics of the 95 confirmed vHAP/VAP episodes

*ESBL-E* extended-spectrum β-lactamase-producing *Enterobacterales, ICU* intensive care unit, *mPCR* multiplex polymerase chain reaction, *PaO<sub>2</sub>/FiO<sub>2</sub>* ratio of the partial pressure of arterial oxygen to the fraction of inspired oxygen, *SOFA* sequential organ failure assessment, *VAP* ventilator associated pneumonia, *vHAP* ventilated hospital-acquired pneumonia

Continuous variables are expressed as median [interquartile range] and compared using Wilcoxon's rank test; categorical variables are expressed as n (%) and compared using Chi-square or Fisher's exact tests, as appropriate. No adjustment for multiple comparisons was performed

<sup>1</sup> Citrobacter Koseri (n = 1), Citrobacter Amalonaticus (n = 1), Klebsiella Aerogenes (n = 1), Klebsiella Oxytoca (n = 2)

<sup>2</sup> Circulatory failure is defined as cardiovascular SOFA score  $\geq$  3

of 7.5 (2.1–35.9), p=0.004), propensity-weighting model (aOR of 5.9 (1.6–22.1), p=0.008), and matching-cohort model (aOR of 5.8 (1.5–22.1), p=0.01), eTable 6. Results were similar in the sensitivity analysis including only the first pneumonia episode (eTable 7). The time required to shift to optimal antibiotic therapy tended to be shorter for patients in the mPCR group, as compared with their counterparts: 9 [3–45] hours vs. 30 [20–55] hours, p=0.09 (Table 3, eFigure 2). Similar results were obtained from the sensitivity analysis conducted on only the first pneumonia episode: 24 [3–45] hours vs. 30 [21–50] hours, p=0.09 (eFigure 2). An exploratory analysis focusing on the first episode of pneumonia (N=59, of which 17 had mPCR testing), found no significant difference in the number of carbapenem treatment days over the seven days following the sampling between mPCR and the conventional groups (2 [0–7] days vs. 2 [0–5] days, P=0.73), even if only ESBL-E non-related cases (N=36, of which 11 had mPCR) were considered (0 [0–2] day vs. 0 [0–1.5] day, P=0.81). Five patients (8.5%) had positive microbiological samples for CRB within the seven days following their first episode of VAP/vHAP: *Stenotrophomonas maltophilia* (protected telescopic catheter N=1, mPCR group; skin culture in a patient with toxic epidermal necrolysis N=1, conventional group), carbapenem-resistant *Pseudomonas aeruginosa* (skin culture in a patient with toxic epidermal necrolysis N=1, conventional group), and *NDM*-producing *Escherichia coli* 

**Table 3** Empirical antibiotic therapy adequation according to the use of mPCR and ESBL-E related pneumonia status in the 95 episodes of nosocomial pneumonia in the mechanically ventilated ESBL-E carriers

| All episodes   | Conventional group, n=73   | mPCR group, n = 22 | р       |
|--|----------------------------|--------------------|---------|
| Empirical antibiotic therapy after sampling                                    |                            |                    |         |
| No initiation  | 24 (33)                    | 6 (27)             | 0.6     |
| Non-carbapenem β-lactam  | 20 (27)                    | 9 (41)             | 0.2     |
| Carbapenem   | 29 (40)                    | 7 (32)             | 0.5     |
| Combination therapy for Gram-negative coverage                                 | 20 (27)                    | 10 (45)            | 0.1     |
| Empirical antibiotic therapy after mPCR result                                 |                            |                    |         |
| No initiation  | 24 (33)                    | 0                  | 0.002   |
| Non-carbapenem β-lactam  | 20 (27)                    | 6 (27)             | 1       |
| Carbapenem   | 29 (40)                    | 16 (73)            | 0.007   |
| Combination therapy for Gram-negative coverage                                 | 20 (27)                    | 7 (32)             | 0.7     |
| Antibiotic therapy adequation  |                            |                    |         |
| Adequate empirical antibiotic therapy (excluding aminoglycosides) <sup>1</sup> | 30 (41)                    | 19 (86)            | < 0.001 |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 31 (42)                    | 19 (86)            | < 0.001 |
| Optimal empirical antibiotic therapy <sup>2</sup>                              | 20 (27)                    | 15 (68)            | 0.001   |
| Time required for optimal antibiotic therapy, hours                            | 30 [20–55]                 | 9 [3–45]           | 0.09    |
| ESBL-E related pneumonia   | Conventional group, n = 38 | mPCR group, n=9    | р       |
| Adequate empirical antibiotic therapy (excluding aminoglycosides) <sup>1</sup> | 13 (34)                    | 9 (100)            | < 0.001 |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 14 (37)                    | 9 (100)            | < 0.001 |
| Optimal empiric antibiotic therapy <sup>2</sup>                                | 13 (34)                    | 9 (100)            | 0.001   |
| Non ESBL-E related pneumonia   | Conventional group, n = 35 | mPCR group, n = 13 | р       |
| Adequate empirical antibiotic therapy (excluding aminoglycosides) <sup>1</sup> | 17 (49)                    | 10 (77)            | 0.08    |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 17 (49)                    | 10 (77)            | 0.08    |
| Optimal empirical antibiotic therapy <sup>2</sup>                              | 7 (20)                     | 6 (46)             | 0.1     |
| Overconsumption of carbapenem <sup>3</sup>                                     | 7 (20)                     | 7 (54)             | 0.03    |
| Ventilator-associated pneumonia  | Conventional group, n = 73 | mPCR group, n = 20 | р       |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 31 (42)                    | 18 (90)            | < 0.001 |
| Patients without carbapenem within 72 h prior to sample                        | Conventional group, n = 56 | mPCR group, n = 22 | р       |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 21 (37)                    | 19 (86)            | < 0.001 |
| First episode of VAP/vHAP  | Conventional group, n=42   | mPCR group, n = 17 | р       |
| Adequate empirical antibiotic therapy (including aminoglycosides) <sup>1</sup> | 17 (40)                    | 15 (88)            | < 0.001 |
|  |                            |                    |         |

ESBL-E extended-spectrum  $\beta$ -lactamase-producing Enterobacterales, mPCR multiplex polymerase chain reaction

Categorical variables are expressed as n (%) and compared using Chi-square or Fisher's exact tests as appropriate. No adjustment for multiple comparisons was performed

<sup>1</sup> Empirical antibiotic therapy was considered adequate if at least one agent was active on all of the offensive pathogens identified by the conventional microbiological culture, based on antibiotic susceptibility findings

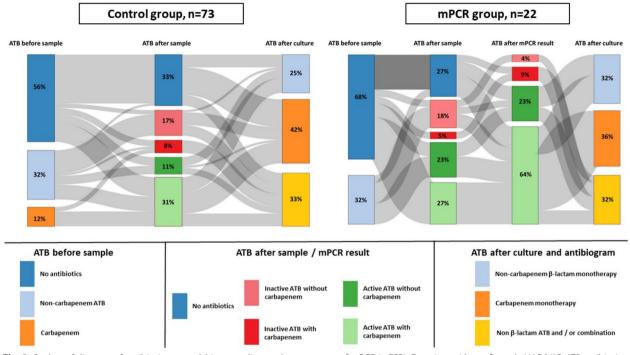
<sup>2</sup> Empirical antibiotic therapy was considered optimal if it was not only active but also not excessively broad-spectrum

<sup>3</sup> Overconsumption of carbapenem was defined as an empirical use of carbapenem whenever the causative bacteria was susceptible to a first-line β-lactam

(protected telescope catheter N=1, mPCR group; urine culture N=1, conventional group).

#### Discussion

To the best of our knowledge, we herein report the first study on mPCR testing specifically focused on ESBL-E carriers, with the following main results: (i) in suspected vHAP/VAP,  $bla_{CTX-M}$  had an excellent concordance with standard culture to rule out ESBL-E-related pneumonia; (ii) in confirmed vHAP/VAP, an mPCR-based approach significantly increased the rate of prescribing adequate and optimal empirical antibiotic therapy in the specific context of our ICU with a restrictive antibiotic policy. AMS for suspected vHAP/VAP in ESBL-E carriers is a



**Fig. 2** Sankey of diagram of antibiotic stewardship according to the use or not of mPCR in ESBL-E carriers with confirmed vHAP/VAP. *ATB* antibiotic therapy, *ESBL-E* extended-spectrum  $\beta$ -lactamase-producing *Enterobacterales*, *mPCR* multiplex polymerase chain reaction, *MV* mechanical ventilation, *VAP* ventilator associated pneumonia, *vHAP* ventilated hospital-acquired pneumonia

daily challenge for intensivists who need to choose the most likely active antibiotic to give in case pneumonia settles [26], and to decide which episodes to treat, since ventilator-associated events mostly reflect non-infectious events [4].

The overall diagnostic value of mPCR we observed is consistent with previous studies findings [6–10]. The reported concordance of negative  $bla_{CTX-M}$  result with the culture helps to eliminate ESBL-E-induced pneumonia and consequently to serenely spare carbapenems upon dealing with suspected vHAP/VAP in ESBL-E carriers. Multicenter studies using Biofire<sup>®</sup> Filmarray<sup>®</sup> also reported a 100% negative concordance of  $bla_{CTX-M}$  to rule out the diagnosis of ESBL-E related pneumonia, but included very few of such cases [7, 8]. mPCR approach is entangled by with two inherent limitations: (i) the risk of false negatives generated by *Enterobacterales* that are not included in the mPCR panel [6]; (ii) its inadequacy in countries where  $bla_{CTX-M}$  is not the predominant gene expressed by ESBL-E.

AMS is a challenging but crucial matter in ICU, especially in ESBL-E carriers. Generalising prescription of carbapenems to ESBL-E carriers is not a suitable approach for several reasons. First, as previously observed, a quarter of pneumonia cases CRB-induced [27]. Second, unnecessary exposure to carbapenems

multiplies the risk of triggering CRB in future infections [28–30]. Third, recent studies described a positive impact of a restrictive antibiotic policy [22, 31]. In our study, confirmed VAP accounted for less than half, and ESBL-E-related VAP for less than a quarter of the suspected pneumonia episodes, which is in line with previous reports [4]. mPCR use could therefore guide decisionmaking process for AMS in ESBL-E carriers, especially when physician decided to initiate antibiotic therapy for whom guidelines recommend the use of carbapenems as empirical antibiotic therapy [3], (i) by enhancing a reasonable restrictive AMS policy that precludes carbapenems facing suspected VAP/vHAP, thanks to the high reported performance value of *bla*<sub>CTX-M</sub> to rule out the diagnosis of ESBL-E related pneumonia; (ii) by increasing the rate of prescribing adequate and optimal empirical antibiotic therapy in confirmed VAP/vHAP. However, if mPCR is used in very low pre-test clinical probability of pneumonia, due to the high sensitivity and the rate of overdiagnosed pneumonia, the risk of overconsumption of carbapenem may prevail. An algorithm for the use of mPCR in ESBL-E carriers with a suspected VAP/ vHAP is proposed in eFigure 3. Nonetheless and given the conflicting results recently reported by randomised controlled trials on mPCR [11, 32], the impact mostly pronounced in the initial hours following respiratory

sampling and the cost of individual tests, the role of mPCR in AMS for ICU patients needs further investigations. Indeed, most studies using mPCR showed no difference in number of days alive and free from antibiotics or the duration of use of broad spectrum antibiotics [11, 12]. A promising area of application could be specific situations, such as patients at risk from MDR bacteria.

Our study has several limitations. First, it is monocentric with a small number of patients, which implies a cautious interpretation of our findings. These results need to be confirmed by large multicentre studies including ICUs with various local ecology and antibiotic policy. Our findings are not applicable in regions with ESBL-E mainly due to TEM- or SHV-producing isolates. Second, the inclusion of multiple episodes related to the same patient might be a source of bias, but results were similar in the sensitivity analysis including only the first pneumonia episode. Third, mPCR was performed at the physician's discretion resulting in an imbalance in some important variables (shock, exposure to carbapenems) between the mPCR and conventional groups. However, we present a real-life picture of an mPCR-based AMS focused on this high-risk ICU population. In addition, the propensity-weighting, the matching-cohort, and the multivariable logistic regression models showed that the mPCR-based approach was independently associated with better antibiotic stewarding towards more adequate and optimal empirical antibiotic therapy. Yet, the use of these models in a small sample needs to be interpreted cautiously. Fourth, in our study, we did not provide data on the cost effectiveness and the ecological impacts of such an approach. These results are preliminary and need to be evaluated in prospective randomised clinical trials. The latter will have to evaluate the ecological impact of a mPCR-based AMS (i.e., antibiotic resistance rates, carbapenems consumption) and the cost-effectiveness of such an approach.

### Conclusion

mPCR  $bla_{CTX-M}$  showed an excellent diagnostic value to rule out the diagnosis of ESBL-E related pneumonia in ESBL-E carriers with suspected VAP/vHAP. The secondary analysis of the use of mPCR in confirmed VAP/vHAP found that a mPCR-based approach was associated with increased prescription of adequate empirical antibiotic therapy. Performing mPCR on respiratory samples seems to be a promising tool in ESBL-E carriers with suspected vHAP/VAP. However, if mPCR is used in very low pretest clinical probability of pneumonia, due to the high sensitivity and the rate of overdiagnosed pneumonia, the risk of overconsumption of carbapenem may prevail. Further studies are warranted.

#### Abbreviations

| ADDICVIC | Abbieviations   |  |  |  |
|----------|---|--|--|--|
| AMS      | Antimicrobial stewardship                                       |  |  |  |
| ARDS     | Acute respiratory distress syndrome                             |  |  |  |
| AST      | Antibiotic susceptibility testing                               |  |  |  |
| CRB      | Carbapenem-resistant bacteria                                   |  |  |  |
| ESBL-E   | Extended-spectrum β-lactamase-producing <i>Enterobacterales</i> |  |  |  |
| FN       | False negative  |  |  |  |
| FP       | False positive  |  |  |  |
| ICU      | Intensive care unit   |  |  |  |
| MDR      | Multidrug-resistant   |  |  |  |
| mPCR     | Multiplex PCR   |  |  |  |
| MV       | Mechanical ventilation  |  |  |  |
| SAPS     | Simplified acute physiology score                               |  |  |  |
| SOFA     | Sequential organ failure assessment                             |  |  |  |
| TN       | True negative   |  |  |  |
| TP       | True positive   |  |  |  |
| VAP      | Ventilator-associated pneumonia                                 |  |  |  |
| vhap     | Ventilated hospital-acquired pneumonia                          |  |  |  |
| CTX-M    | Cefotaximase-Munich   |  |  |  |
| NPA      | Negative percentage agreement                                   |  |  |  |
| PPA      | Positive percentage agreement                                   |  |  |  |
|          |   |  |  |  |

#### Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13613-024-01348-5.

#### Supplementary Material 1.

Supplementary Material 2.

Supplementary Material 3. eFigure 1. Propensity score balance. Comparisons of the absolute standardised mean differences on selected covariates (circulatory failure defined as cardiovascular SOFA score  $\geq$  3, PaO2/FiO2 < 150 mmHg and the use of carbapenem within 72 h prior to sample), before and after weighting and matching.

Supplementary Material 4. eFigure 2. Kaplan-Meier curve of the proportion of patients receiving optimal antibiotic therapy according to whether mPCR was used or not (censoring threshold: 48 h). A. Kaplan-Meier Curve for optimal antibiotic therapy according to the use of mPCR in the whole cohort (N=95). P-value was determined using the log-rank test. B. Kaplan-Meier Curve for optimal antibiotic therapy according to the use of mPCR in the group of the term of the set of the use of mPCR in the first episode of pneumonia (N=59). P-value was determined using the log-rank test.

Supplementary Material 5. eFigure 3. Proposed Algorithm for empiric antibiotic therapy in ESBL-E carriers with a suspicion of VAP. CTX-M, Cefo-taximase-Munich; ESBL-E, extended-spectrum  $\beta$ -lactamase-producing *Enterobacterales*; mPCR, multiplex polymerase chain reaction; VAP, ventilator associated pneumonia.

Supplementary Material 6.

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None.

#### Author contributions

PB, KR, VF and PLW designed the study and analysed the data. PB, KR and VF were responsible for data acquisition. PB, VF, PLW, BP, SG, RA, PL, EM, AG, GC, NdP, AMD and KR contributed to the study design and analysis, interpretation of data, drafting of initial manuscript, critical revision of intellectual content, and approval of the submitted version of the article. PB and KR are the guarantor of study data integrity. PB and KR had full access to all of the study data and are deemed responsible for data integrity and accuracy of their analyses.

#### Funding

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#### Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

#### Declarations

#### Ethics approval and consent to participate

This observational study was approved by the Institutional Review Board of Henri Mondor university hospital Patients were informed of their inclusion in the study and informed consent was waived as per French law. The database was registered, number 20181128165057.

#### Consent for publication

Not applicable.

#### **Competing interests**

KR reports personal fees from Shionogi and MSD. GC reports personal fees from Air Liquide Medical System, GE Healthcare, Dräger, Fisher and Paykel, Medtronic and Löwenstein, outside the submitted work. AMD reports grants from Fischer Paykel, Baxter, Philips, Ferring and GSK, personal fees from Air Liquide, Baxter, Amomed, Getingue and Addmedica, outside the submitted work. All other authors declare that they have no competing interests.

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#### References

- 1. De Bus L, Gadeyne B, Steen J, Boelens J, Claeys G, Benoit D, et al. A complete and multifaceted overview of antibiotic use and infection diagnosis in the intensive care unit: results from a prospective four-year registration. Crit Care Lond Engl. 2018;22(1):241.
- Detsis M, Karanika S, Mylonakis E. ICU acquisition rate, risk factors, and clinical significance of digestive tract colonization with extended-spectrum beta-lactamase-producing enterobacteriaceae: a systematic review and meta-analysis. Crit Care Med. 2017;45(4):705–14.
- Haute Autorité de Santé. Antibiothérapie des infections à entérobactéries et à Pseudomonas aeruginosa chez l'adulte : place des carbapénèmes et de leurs alternatives. https://www.has-sante.fr/jcms/c\_2968915/fr/antib iotherapie-des-infections-a-enterobacteries-et-a-pseudomonas-aerug inosa-chez-l-adulte-place-des-carbapenemes-et-de-leurs-alternatives. Accessed 16 Sep 2022.
- Barbier F, Bailly S, Schwebel C, Papazian L, Azoulay É, Kallel H, et al. Infection-related ventilator-associated complications in ICU patients colonised with extended-spectrum β-lactamase-producing Enterobacteriaceae. Intensive Care Med. 2018;44(5):616–26.
- 5. Bush K, Bradford PA. Epidemiology of  $\beta$ -lactamase-producing pathogens. Clin Microbiol Rev. 2020;33(2):e00047-e119.
- Gastli N, Loubinoux J, Daragon M, Lavigne JP, Saint-Sardos P, Pailhoriès H, et al. Multicentric evaluation of BioFire FilmArray Pneumonia Panel for rapid bacteriological documentation of pneumonia. Clin Microbiol Infect. 2021;27(9):1308–14.
- Molina FJ, Botero LE, Isaza JP, Cano LE, López L, Tamayo L, et al. Diagnostic concordance between BioFire<sup>®</sup> FilmArray<sup>®</sup> Pneumonia Panel and culture in patients with COVID-19 pneumonia admitted to intensive care units: the experience of the third wave in eight hospitals in Colombia. Crit Care Lond Engl. 2022;26(1):130.
- Maataoui N, Chemali L, Patrier J, Tran Dinh A, Le Fèvre L, Lortat-Jacob B, et al. Impact of rapid multiplex PCR on management of antibiotic therapy

in COVID-19-positive patients hospitalized in intensive care unit. Eur J Clin Microbiol Infect Dis. 2021;40(10):2227–34.

- Razazi K, Delamaire F, Fihman V, Boujelben MA, Mongardon N, Gendreau S, et al. Potential of multiplex polymerase chain reaction performed on protected telescope catheter samples for early adaptation of antimicrobial therapy in ARDS patients. J Clin Med. 2022;11(15):4366.
- Enne VI, Aydin A, Baldan R, Owen DR, Richardson H, Ricciardi F, et al. Multicentre evaluation of two multiplex PCR platforms for the rapid microbiological investigation of nosocomial pneumonia in UK ICUs: the INHALE WP1 study. Thorax. 2022;77(12):1220–8.
- Fartoukh M, Nseir S, Mégarbane B, Cohen Y, Lafarge A, Contou D, et al. Respiratory multiplex PCR and procalcitonin to reduce antibiotic exposure in severe SARS-CoV-2 pneumonia: a multicentre randomized controlled trial. Clin Microbiol Infect. 2023;S1198-743X(23)00031–9.
- Markussen DL, Serigstad S, Ritz C, Knoop ST, Ebbesen MH, Faurholt-Jepsen D, et al. Diagnostic stewardship in community-acquired pneumonia with syndromic molecular testing: a randomized clinical trial. JAMA Netw Open. 2024;7(3): e240830.
- Plachouras D, Lepape A, Suetens C. ECDC definitions and methods for the surveillance of healthcare-associated infections in intensive care units. Intensive Care Med. 2018;44(12):2216–8.
- Martin-Loeches I, Reyes LF, Nseir S, Ranzani O, Povoa P, Diaz E, et al. European Network for ICU-Related Respiratory Infections (ENIRRIs): a multinational, prospective, cohort study of nosocomial LRTI. Intensive Care Med. 2023;49(10):1212–22.
- Peiffer-Smadja N, Bouadma L, Mathy V, Allouche K, Patrier J, Reboul M, et al. Performance and impact of a multiplex PCR in ICU patients with ventilator-associated pneumonia or ventilated hospital-acquired pneumonia. Crit Care Lond Engl. 2020;24(1):366.
- Kollef MH, Nováček M, Kivistik Ü, Réa-Neto Á, Shime N, Martin-Loeches I, et al. Ceftolozane-tazobactam versus meropenem for treatment of nosocomial pneumonia (ASPECT-NP): a randomised, controlled, double-blind, phase 3, non-inferiority trial. Lancet Infect Dis. 2019;19(12):1299–311.
- Jarlier V, Nicolas MH, Fournier G, Philippon A. Extended broad-spectrum beta-lactamases conferring transferable resistance to newer beta-lactam agents in Enterobacteriaceae: hospital prevalence and susceptibility patterns. Rev Infect Dis. 1988;10(4):867–78.
- Poritz MA, Blaschke AJ, Byington CL, Meyers L, Nilsson K, Jones DE, et al. FilmArray, an automated nested multiplex PCR system for multipathogen detection: development and application to respiratory tract infection. PLoS ONE. 2011;6(10): e26047.
- Edin A, Eilers H, Allard A. Evaluation of the Biofire Filmarray Pneumonia panel plus for lower respiratory tract infections. Infect Dis Lond Engl. 2020;52(7):479–88.
- 20. Poole S, Clark TW. Rapid syndromic molecular testing in pneumonia: the current landscape and future potential. J Infect. 2020;80(1):1–7.
- Health C for D and R. Statistical guidance on reporting results from studies evaluating diagnostic tests—guidance for industry and FDA staff. FDA; 2020. https://www.fda.gov/regulatory-information/search-fda-guida nce-documents/statistical-guidance-reporting-results-studies-evalu ating-diagnostic-tests-guidance-industry-and-fda. Accessed 3 Apr 2024.
- Hranjec T, Rosenberger LH, Swenson B, Metzger R, Flohr TR, Politano AD, et al. Aggressive versus conservative initiation of antimicrobial treatment in critically ill surgical patients with suspected intensive-care-unitacquired infection: a quasi-experimental, before and after observational cohort study. Lancet Infect Dis. 2012;12(10):774–80.
- Kalil AC, Metersky ML, Klompas M, Muscedere J, Sweeney DA, Palmer LB, et al. Management of adults with hospital-acquired and ventilatorassociated pneumonia: 2016 Clinical Practice Guidelines by the Infectious Diseases Society of America and the American Thoracic Society. Clin Infect Dis. 2016;63(5):e61-111.
- 24. Weiss E, Zahar JR, Lesprit P, Ruppe E, Leone M, Chastre J, et al. Elaboration of a consensual definition of de-escalation allowing a ranking of  $\beta$ -lactams. Clin Microbiol Infect. 2015;21(7):649.e1-10.
- von Elm E, Altman DG, Egger M, Pocock SJ, Gøtzsche PC, Vandenbroucke JP. The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. Lancet. 2007;370(9596):1453–7.
- Zaragoza R, Vidal-Cortés P, Aguilar G, Borges M, Diaz E, Ferrer R, et al. Update of the treatment of nosocomial pneumonia in the ICU. Crit Care Lond Engl. 2020;24(1):383.

- Razazi K, Mekontso Dessap A, Carteaux G, Jansen C, Decousser JW, de Prost N, et al. Frequency, associated factors and outcome of multidrug-resistant intensive care unit-acquired pneumonia among patients colonized with extended-spectrum β-lactamase-producing Enterobacteriaceae. Ann Intensive Care. 2017;12(7):61.
- Armand-Lefèvre L, Angebault C, Barbier F, Hamelet E, Defrance G, Ruppé E, et al. Emergence of imipenem-resistant gram-negative bacilli in intestinal flora of intensive care patients. Antimicrob Agents Chemother. 2013;57(3):1488–95.
- Luyt CE, Aubry A, Lu Q, Micaelo M, Bréchot N, Brossier F, et al. Imipenem, meropenem, or doripenem to treat patients with Pseudomonas aeruginosa ventilator-associated pneumonia. Antimicrob Agents Chemother. 2014;58(3):1372–80.
- McLaughlin M, Advincula MR, Malczynski M, Qi C, Bolon M, Scheetz MH. Correlations of antibiotic use and carbapenem resistance in enterobacteriaceae. Antimicrob Agents Chemother. 2013;57(10):5131–3.
- 31. Le Terrier C, Vinetti M, Bonjean P, Richard R, Jarrige B, Pons B, et al. Impact of a restrictive antibiotic policy on the acquisition of extended-spectrum beta-lactamase-producing Enterobacteriaceae in an endemic region: a before-and-after, propensity-matched cohort study in a Caribbean intensive care unit. Crit Care Lond Engl. 2021;25(1):261.
- Darie AM, Khanna N, Jahn K, Osthoff M, Bassetti S, Osthoff M, et al. Fast multiplex bacterial PCR of bronchoalveolar lavage for antibiotic stewardship in hospitalised patients with pneumonia at risk of Gram-negative bacterial infection (Flagship II): a multicentre, randomised controlled trial. Lancet Respir Med. 2022;10(9):877–87.

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