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# Nutritional and metabolic modulation of inflammation in critically ill patients: a narrative review of rationale, evidence and grey areas

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# **Abstract**

**Background** Inflammation is the hallmark of critical illness and triggers the neuro-endocrine stress response and an oxidative stress. Acute inflammation is initially essential for patient's survival. However, ongoing or exaggerated inflammation, due to persistent organ dysfunction, immune dysfunction or poor inflammation resolution, is associated to subsequent hypermetabolism and hypercatabolism that severely impact short and long-term functional status, autonomy, as well as health-related costs. Modulation of inflammation is thus tempting, with the goal to improve the short- and long-term outcomes of critically ill patients.

**Findings** Inflammation can be modulated by nutritional strategies (including the timing of enteral nutrition initiation, the provision of some specific macronutrients or micronutrients, the use of probiotics) and metabolic treatments. The most interesting strategies seem to be n-3 polyunsaturated fatty acids, vitamin D, antioxidant micronutrients and propranolol, given their safety, their accessibility for clinical use, and their benefits in clinical studies in the specific context of critical care. However, the optimal doses, timing and route of administration are still unknown for most of them. Furthermore, their use in the recovery phase is not well studied and defined.

**Conclusion** The rationale to use strategies of inflammation modulation is obvious, based on critical illness pathophysiology and based on the increasingly described effects of some nutritional and pharmacological strategies. Regretfully, there isn't always substantial proof from clinical research regarding the positive impacts directly brought about by inflammation modulation. Some arguments come from studies performed in severe burn patients, but such results should be transposed to non-burn patients with caution. Further studies are needed to explore how the modulation of inflammation can improve the long-term outcomes after a critical illness.

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OR, USA

**Keywords** Inflammation, Critical illness, Intensive care, Burn injury, Nutrition, Metabolism, Stress response, Pharmaconutrition

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# **Background**

A common feature in all critically ill patients is the systemic inflammation that occurs in response to tissue damage (due to infection, trauma or surgery) and proinflammatory mediators. The inflammatory response to tissue damage is induced by a complex array of mediators being interrelated for large part and involves the activation of various cells such as leukocytes and other inflammatory cells, leading to a massive production of reactive oxygen species (ROS). Inflammation initiates a protective response for the destruction of pathogens and serves as chemical attractants for beginning the wound repair. Systemic inflammation further takes the form of a continuum of severity, with potential fluctuations according to the successive hits (for example surgeries, infections). At a cellular level, an impaired mitochondrial function is noted, limiting its metabolic capacities. Inflammation as well triggers a neuro-endocrine stress response, characterized by an adrenergic stimulation, an elevation in cortisol level, a blunted activity of the somatotropic axis and a hypogonadism  $[1]$  $[1]$ . In case of ongoing inflammation, the neuroendocrine stress response leads to supraphysiologic metabolic rates (hypermetabolism), protein hypercatabolism (in muscles and bones), insulin resistance, alterations in lipid metabolism and fat composition. Catecholamines are a main effector for hypermetabolism, in link with the browning of white adipose tissue [[2\]](#page-7-1). These metabolic changes are initially essential for patient's survival, ultimately providing the brain and the immune system with fuel. However, vicious cycles can develop, further potentiating systemic inflammation. Hypercatabolism is associated with immune dysfunction, increasing the susceptibility to recurrent sepsis. Persisting organ injuries, in part due to oxidative stress, trigger recurrent alarmins release: mitochondrial danger-associated molecular patterns (DAMPs) are released from injured or necrotic cells and enhance the inflammatory reaction [[3\]](#page-7-2). As example, mitochondrial oxidative damages have been observed during prolonged mechanical ventilation and diaphragmatic inactivity [\[4](#page-7-3)]. The hypothesis of a poor resolution of inflammation, due to defects in pro-resolving pathways, could also participate in chronic inflammation, influenced by age, lifestyle factors, epigenetic or microbiome [\[5\]](#page-7-4). A chronic hyperinflammatory hypermetabolic state is a characteristic of chronic critical illness [[6](#page-7-5)]. The definition of what is considered a continuous inflammation and a chronic critical illness is not well defined, is influenced by preadmission clinical conditions and by the severity of the critical illness, but is probably measured in days [[7,](#page-7-6) [8](#page-7-7)].

A clinically meaningful example of such condition is the severe burn injury (traditionally defined as burns affecting more than 20% of the total body surface area in adults  $[9]$  $[9]$ ). In these patients, the metabolic derangements are the most intense compared to those encountered in other critical conditions such as sepsis and trauma, and are known to persist for up to years following injury [\[10](#page-7-9)]. However, alike observations have been made in non-burn critically ill patients. After COVID-19 acute respiratory distress syndrome (ARDS), a progressive hypermetabolism has been demonstrated during the ICU stay through the use of indirect calorimetry  $[11]$  $[11]$ . Signs of persistent inflammation have also been observed 3 months after discharge from intensive care unit (ICU), in association with persistent hypermetabolism and impaired exercise capacity [\[12,](#page-8-0) [13\]](#page-8-1). Similar findings were described in non-COVID ICU survivors [\[14](#page-8-2)]. Muscle alterations are attributed partly to atrophy, mitochondrial dysfunction and endoplasmic reticulum stress [[15](#page-8-3), [16\]](#page-8-4).

The clinical translation of persistent inflammation and subsequent hypermetabolism is significant: functional status, autonomy, return to work or pre-admission level of activities, health-related costs may be severely impacted. These altered outcomes are part of the postintensive care syndrome [\[17](#page-8-5)]. According to a recent systematic review, long-term outcomes following critical illness are potentially associated with an inflammatory process [\[18\]](#page-8-6). Trying to modulate persistent inflammation does thus make sense. However, it is wise to recognize that inflammation modulation is still a concept, as the boundary between a beneficial and a detrimental level of systemic inflammation is unknown, and probably specific to a given patient in a given clinical situation, depending on the time point in the illness trajectory. Modifying the inflammation level could aim to reduce the overspill of acute inflammation and/or to prevent further exacerbation and/or to help resolving the response, depending on the considered timing.

Inflammation modulation may be based on nutritional interventions and pharmacological modalities. Nutritional interventions refer to an optimal timing and route of feeding, while pharmaco-nutrition refers to the addition of supraphysiological doses of specific macro- and micronutrients to standard feeding, relying on their ability of acting on inflammatory and immune pathways whatever the nutritional status of the patient [[19](#page-8-7)]. These beneficial effects are dissociated from their nutritional properties, in terms of energy supply for example [\[20](#page-8-8)].

The strategies of inflammation modulation that have, according to the current literature, a prominent role while being commercially available in daily practice are presented and discussed in the present narrative review (Table [1](#page-2-0)). Their potential effects on the subsequent hypermetabolism and hypercatabolism are also described. It should be noted that the presented list may be biased by a subjective selection and omits new or revised drugs that are still being studied in preclinical models [[21\]](#page-8-9).



<span id="page-2-0"></span>**Table 1** Recommended, suggested, possible and banned strategies to modulate hyperinflammation in critically ill patients and survivors:

NR: not recommended (signal of harm or absence of benefit); OI: of interest (could be considered in view of the rationale and the available literature, but evidence and practical recommendations still lacking); R: recommended in guidelines; S: suggested in guidelines; TBE: to be explored (real rationale but lack of available literature)

References of available literature and guidelines: refer to the text

Studies in both severe burn patients and non-burn critically ill patients were examined. Severe burn injury is the most severe form of trauma or critical illness in terms of the debilitating stress response it provokes [\[22](#page-8-10)], and is thus recognized as a model of systemic inflammation following injury. The hypermetabolic response in severe burn patients has been largely studied, as well as the manipulation of this response, especially because the injury is easy to quantify, as the percentage of body surface area burned is measured objectively. However, exudative losses of proteins and micronutrients make this model particular. The stress response in severe burn patients is also much more intense and prolonged than in trauma or septic patients [\[23\]](#page-8-11). Results observed in studies performed in severe burn patients should not be simply transposed to non-burn critically ill patients but are obviously food for thought.

# **Clinical assessment of inflammation in critically ill patients** Inflammation monitoring relies on blood biomarkers, mainly acute phase proteins (essentially C-reactive protein (CRP), fibrinogen, serum amyloid A or procalcitonin), cytokines (mainly interleukines (IL) 6 and 10 or TNF $\alpha$ ), or immune cells degranulation markers (such as myeloperoxidase). Not all of these markers are readily

Conflicting observations mitigate the potential interest of CRP to detect persistent inflammation: CRP remains elevated in burn patients several months after injury [\[24](#page-8-12)], while it has been shown to decrease more rapidly than

available in every lab for daily use at bedside.

other inflammatory markers in other populations of ICU survivors [[12,](#page-8-0) [13](#page-8-1), [25\]](#page-8-13).

Closed monitoring of the inflammation status during the entire ICU and post-ICU trajectory is thus difficult, making individualization of its modulation approximate, if not based on surrogate markers such as body composition or measured energy expenditure.

# **Inflammation modulation using medical nutrition**

The primary goal of medical nutrition is to provide an adequate supply in nutrients to maintain organ function, to minimize the devastating effects of persistent catabolism and to optimize healing and immune function.

When focusing on inflammation, the timing of medical nutrition initiation could matter, especially when considering enteral route. In severe burn patients, early institution of trophic enteral feeding (i.e. within the first 12 to 24 h after injury) can significantly reduce the level of inflammation, based on plasma C-reactive protein (CRP) levels [[26](#page-8-14)] and can prevent further total energy deficit [[27\]](#page-8-15), although the effects of early initiation on hypermetabolism are debated [\[28](#page-8-16)]. In non-burn ICU patients, the impact of feeding initiation timing on inflammation markers is still underrecognized, despite the fact that there has been much discussion on the optimal time to provide full nutrition.

The provision of the energy needs after the first days following ICU admission, could also matter. When enable to provide the defined nutritional targets by enteral route, using supplemental parenteral nutrition has been

associated with reduced serum levels of pro-inflammatory cytokines and a decrease in CRP over the next 5 days, compared to enteral nutrition alone [[29\]](#page-8-19).

Altogether, it can be assumed that early enteral nutrition is a way to preserve gut integrity, subsequently reducing secondary infections, and also reducing toxic bacterial metabolites while enhancing production of short chain fatty acid with local and systemic effects [\[30](#page-8-20)]. However, in absence of strong evidence, these effects remain speculative and should be further studied.

Inflammation is known to induce insulin resistance, gastro-intestinal dysfunction and anorexia, essentially limiting the entry of nutrients into cells. In a recent secondary analysis of the EFFORT cohort, including inhospital patients (outside ICU) at risk of malnutrition, patients with high inflammation (as reflected by CRP levels) tended to benefit less from nutritional support with regards to 30-day mortality, compared to patients with less inflammation [[31\]](#page-8-21). However, it is still unknown if the timing of nutritional support initiation in ICU patients should be individualized according to the inflammation level.

# **Inflammation modulation using specific macronutrients** *Glutamine*

Glutamine is the preferred oxidative fuel for rapidly divide cells such as lymphocytes and gut mucosal cells. Glutamine is considered a conditionally essential amino acid: in case of metabolic stress, peripheral demand outstrips the production, leading to depletion. Glutamine is able to attenuate the inflammatory response via effects on heat shock protein, nuclear factor- **κ**B signaling pathway and via the attenuation of tumor necrosis factor- $\alpha$ , IL-6, and IL-18 expression after sepsis [\[32\]](#page-8-22).

The signal coming from a number of small randomized studies investigating glutamine supplementation in severe burn patients were positive, with a reduction in hospital stay and mortality, as well as gram-negative bacteremia [\[33](#page-8-23)]. Another small study also demonstrated that glutamine supplementation could reduce the resting energy expenditure and the catecholamine blood levels in severe burn adults [\[34\]](#page-8-24). However, recently, the results of the RE-ENERGIZE multicenter randomized study failed to demonstrate any effect of a 0.5 g/kg/day dose of enteral glutamine on time-to-discharge alive, mortality or gramnegative bacteremia [[35](#page-8-25)]. Importantly glutamine administration was not associated with adverse events. On the contrary, this pragmatic large study included heterogenous patients regarding burn surface area, with very different anticipated exudative glutamine loss [\[36](#page-8-26)]. Up to now, in the latest published guidelines on clinical nutrition in ICU, glutamine administration is still suggested in case of major burns [\[37](#page-8-18)].

Enteral glutamine administration is also recommended in trauma patients or in patients with complicated wound healing [\[37](#page-8-18), [38\]](#page-8-17). In other critically ill patients, signal of harm came from the REDOXS study [\[39\]](#page-8-27), further confirmed by a post-hoc analysis [[40\]](#page-8-28), that demonstrated an increased mortality rate in patients with multiple organ failure receiving high doses of intravenous glutamine. The exact mechanism of injury is unknown, possibly related to the accumulation of glutamine or metabolites in patients with renal dysfunction. Considering this alert, as well as negative meta-analysis [[41\]](#page-8-29), there is no longer a recommendation for an indiscriminate enteral glutamine supplementation during critical illness [\[42](#page-8-30)]. However, an individualized approach with glutamine supplementation in patients with low serum glutamine level or on prolonged parenteral nutrition has never been tested.

#### *Arginine*

Arginine is required for T lymphocyte functions such as proliferation and the expression of normal T cell receptors. It serves to regulate the appropriate immune response to a catabolic challenge and in vasoregulation via inducible nitric oxide synthase (iNOS), a potent vasodilator. Arginine has been found to improve protein kinetics to wound healing, restore T cell function, and promote the transition from the proinflammatory M1 macrophage to an M2 resolution macrophage [[43\]](#page-8-31). Arginine also appears to support the maturation of myeloid derived suppressor cells (MDSC). MDSCs are a heterogeneous group of immature immune cells that originate in bone marrow that can alter and regulate the immune response to an immune challenge. They are characterized by the increased production of reactive oxygen and nitrogen species, and by increased arginase 1 activity [\[44](#page-8-32)]. In excess, MDSCs suppress innate and adaptive immunity and be a major obstacle to a well-orchestrated immune response to challenge. Arginine can enhance the maturation of MDSCs thereby regulating for the appropriate immune response.

L-Arginine supplemented formulations have been in use for greater than four decades with the concept that correction of the relative arginine deficiency noted in critically ill patients would improve systemic immune function. Multiple studies in which supplemental arginine has been delivered in the perioperative period, reported improved clinical outcomes including decreased rates of infections and decreased length of ICU stay [[45–](#page-8-33) [47\]](#page-8-34). Other studies, not all with a strong methodology, also suggested that arginine, supplied enterally at doses available in commercial formulations, may be beneficial to septic patients  $[48]$  $[48]$ . In severe burn patients, arginine supplementation has been rarely studied and is not routinely administered as single agent.

Previously a controversy associated with supplemental arginine in critically ill septic patients was discussed. The controversy revolved around the theoretical concept that supplemental arginine would lead to increase nitric oxide via the upregulation of iNOS and result in refractory hypotension. This theory has been now primarily dismissed following multiple human studies. Reports of infusion of arginine in patients with sepsis or septic shock showed no detrimental hemodynamic effects [\[49](#page-8-36), [50\]](#page-8-37).

Altogether, due to weak evidence of clinical benefits, arginine supplementation for all ICU populations is not recommended.

Citrulline is converted to L-arginine through the activity of argininosuccinate synthetase and argininosuccinate lyase. Citrulline has been described as more efficient than L-arginine at increasing plasma arginine  $[51]$  $[51]$  $[51]$ . A recent study of enteral citrulline supplementation in mechanically ventilated ICU patients confirmed this description, without demonstrating a subsequent beneficial effect on severity scores or immune biomarkers [[52\]](#page-8-39).

### *Carnitine*

L-carnitine, a quaternary ammonium, is synthesized endogenously, and for a largest part, ingested via animalbased food. Its main function is the acyl-carnitine carrier transport system of long-chain fatty acids into mitochondria, where they will undergo subsequent β-oxidation.

Carnitine deficiency, defined as a state of tissue or blood carnitine concentration below the requirement for normal organ function, can lead for instance to muscle weakness. Critically ill patients are at risk of carnitine deficiency, due to clinical conditions, supports or treatments, such as prolonged undernutrition, prolonged total parenteral nutrition (commercial formulations do not contain carnitine), prolonged continuous renal replacement therapy (loss of carnitine in the effluent) or valproate treatment (carnitine used for urinary excretion of drug derivates) [[53](#page-8-40), [54](#page-8-41)]. According to some rare publications on this topic, carnitine deficiency seems scarce in survivors after ICU discharge, while affecting one third of the survivors 3 months after discharge [\[55](#page-8-42)[–57\]](#page-8-43).

Clinical cases of carnitine supplementation have been reported in ICU deficient patients during the acute phase [[53\]](#page-8-40). Interventional trials are rare. In a study including patients whatever their carnitine status, a daily 3 g L-carnitine supplementation during ICU stay led to reduced inflammation markers at the end of intervention compared to control group [[58\]](#page-8-44). Same conclusions were drawn in a more recent similar study in septic patients [\[59\]](#page-8-45). To date, no study investigated the effects of a carnitine supplementation in ICU survivors. However, there are some indications that L-carnitine supplementation could help reducing some blood biomarkers of inflammation and oxidative stress in various categories of non-critically ill patients [\[60](#page-8-46)]. Moreover, growing evidence suggests that L-carnitine could affect muscle mass and function in older adults, probably through a modulation of the muscle protein catabolism / anabolism balance [[61\]](#page-8-47).

# *Omega-3 polyunsaturated fatty acids*

Omega-3 polyunsaturated fatty acids (n-3 PUFA) found in oily fishes and fish oil supplements have anti-inflammatory effects. The fish oils (EPA and DHA) as well as arachidonic acid are precursors of lipoxins, resolvins, maresins and protectins, these endogenously produced compounds act on several cell lines, including monocytes, macrophages, neutrophils, endothelial and dendritic cells. Collectively these are now referred to as Specialized Proresolving Mediators (SPMs). They can inhibit leucocyte chemotaxis, reduce adhesion between leucocytes and endothelium, decrease the production of eicosanoids from arachidonic acid, and decrease the production of inflammatory cytokines [[62\]](#page-8-48). These effects are translated into improvement of clinical outcomes such as ventilation duration, ICU length of stay and infections when n-3 PUFA are given as parenteral emulsion in parallel with a medical nutrition [\[63\]](#page-8-49). Similar positive results have been described with enteral n-3 PUFA [\[64](#page-8-50)]: improvement of oxygenation and length of mechanical ventilation in patients with ARDS [[65](#page-9-1)[–67](#page-9-2)], lower mortality in patients with sepsis [\[68](#page-9-3), [69](#page-9-4)]. SPMs have also been shown to decrease postoperative and neuropathic pain, accelerate removal of inflammatory and necrotic debris, and enhance the conversion from the proinflammatory M1 macrophage to a M2 or resolution macrophage [\[70](#page-9-5)].

Independently of their anti-inflammatory effects, n-3 PUFAs have anabolic effect in muscles, at least partially mediated via the activation of the mTOR pathway [[71\]](#page-9-6). A recent meta-analysis of human studies in heterogeneous population groups identified that lean mass and skeletal muscle mass are favored by higher intakes of EPA and DHA [\[72\]](#page-9-7). Finally, another interesting metabolic perspective comes from an old study in severe burn adults: when receiving a low-fat diet enriched with n-3 PUFA, patients had significantly higher insulin-like growth factor-1 (IGF-1) one month after injury compared to patients receiving high-fat diet or low-fat diet without n-3 PUFA [[73\]](#page-9-8). However, the functional consequence of such endocrine modulation is still to be determined.

Altogether, the use of n-3 PUFAs during the critical care trajectory is relevant and attractive, but future research needs to focus on the best dosages of omega-3 PUFAs to use, the optimal route(s) of administration, the optimal timing and which patients are most likely to benefit [\[74](#page-9-9)].

# **Inflammation modulation using micronutrients** *Micronutrients with direct anti-inflammatory effects*

Vitamin D has a wide range of action, including the classical bone mineralization and remodeling [\[75](#page-9-10)]. Vitamin D has also anti-inflammatory properties, inhibiting the activation of nuclear factor-kappa B (NF-κB) and the production of pro-inflammatory cytokines, such as IL-6 and TNF- $\alpha$  [[76\]](#page-9-11). Some human data suggest that vitamin D could regulate mitochondrial function, especially in muscle, with potential benefits on muscle strength or recovery after exercises [[77\]](#page-9-12). Translation of these findings in the critical care context is currently lacking. From a general point of view, extra-skeletal effects of vitamin D remain debated, mainly due to methodological concerns in the available literature [\[78\]](#page-9-13).

In non-critically ill patients, vitamin D supplementation had significant effects on inflammation and oxidative stress [[79,](#page-9-14) [80\]](#page-9-15). Such effects have been observed in a small study including deficient patients with sepsis who received high dose boluses of cholecalciferol [\[81](#page-9-16)]. However, to date, it is still needed to verify whether the effects of vitamin D on inflammation and mitochondria, through a repletion or a supplementation strategy, modify the short and long-term outcomes of ICU patients or survivors.

Unfortunately, the optimal vitamin D prescription in the critical care context is undetermined, whether in terms of dose, route or timing.

Due to a very high prevalence of hypovitaminosis D in critically ill patients [\[82](#page-9-17)] (and even more in severe burn patients [\[83](#page-9-18)]),it is largely recommended to complete or replete in vitamin D3 to reach a normal vitamin D status [[84\]](#page-9-19). The oral or enteral route is usually used for this purpose. However, considering the gut oedema and the multiple organ dysfunction observed in critically ill patients, it is unclear if the inactive cholecalciferol is adequately absorbed by the gastro-intestinal tract and further converted in its biological active form. These physiological derangements could be the rationale for a more pronounced effect of vitamin D administration on overall mortality or ICU length of stay when given by parenteral route [\[85](#page-9-20)]. Patients presenting a severe vitamin D deficiency could be those who would benefit the most from a vitamin D repletion or supplementation [[86](#page-9-21), [87](#page-9-22)]. It is now increasingly recognized from studies in other medical conditions that ultra-high bolus doses of vitamin D (>100,000 IU) is ineffective due to the subsequent long term activation of the inactivating 24-hydroxylase and fibroblast growth factor 23 [\[88\]](#page-9-23).

# *Micronutrients with antioxidant effects*

Oxidative stress is defined as a state in which the level of toxic ROS overcome the endogenous antioxidant defenses of the host, and damage biologically relevant molecules, leading to mitochondrial damages and systemic inflammation acceleration. Copper, manganese, zinc, iron and selenium are required for the enzymatic endogenous antioxidant defense, while the non-enzymatic mechanisms include vitamins A, C, E [[89](#page-9-24)].

Exogenous antioxidants have been considered for years as part of the ICU nutritional management in attempt to limit the oxidative stress associated critical illness and to improve outcomes such as mortality or illness severity [\[90,](#page-9-25) [91](#page-9-26)]. Despite active research, study results are often conflicting. The variability of the inflammatory response, the oxidative stress, and the endogenous immune response combined with the heterogeneity of the ICU population (in terms of critical condition and amount of exogenous antioxidants required to restore the antioxidant capacity) makes the dosing judgement of antioxidants unclear. The issue of timing of antioxidant administration is also a key factor, as well as the ideal combination of antioxidants [\[92](#page-9-27)]. These are challenging questions to respond to, particularly given that laboratory tests of micronutrient status are an area prone to misuse and misinterpretation [\[93](#page-9-28)].

Vitamin C has been extensively studied in different critical care settings over the past years. In severe burn injury, it has been demonstrated more than 30 years ago that high intravenous doses of vitamin C in the early phase following injury could reduce fluid requirements and burn edema [[94,](#page-9-29) [95](#page-9-30)]. The ongoing VICToRY Trial aims to bring a higher level of evidence regarding this sparing strategy (ClinicalTrials.gov Identifier: NCT04138394). According to two recent meta-analysis in non burn critically ill patients, high dose intravenous vitamin C monotherapy could be associated with a trend toward reduction in overall mortality [[96,](#page-9-31) [97\]](#page-9-32). Interestingly, a high-dose antioxidant supplementation protocol including vitamin C, administered to surgical and trauma patients, led to a reduction of the inflammatory response [[98\]](#page-9-33). Used alone, in a small cohort of patients with severe sepsis, intravenous high dose of vitamin C significantly reduced inflammatory biomarkers [[99](#page-9-34)]. The safety of high doses of vitamin C is still questioned, especially regarding nephropathy and hemolytic anemia in ignored cases of Glucose-6-Phosphate Dehydrogenase deficiency: data providing from published studies are conflicting [[100\]](#page-9-35).

Some weak trends of an inflammation modulation by selenium supplementation in monotherapy has been observed in small cohorts of critically patients at high risk of bias [\[101](#page-9-36)]. Dose and timing of selenium supplementation is unclear, especially since selenium has prooxidant properties which can lead to the opposite of the desired effect.

The use of other antioxidant micronutrients, with the aim at modulating inflammation during or after ICU

stay, has unfortunately not been widely studied in clinical practice.

Overall, at the present time, the guidelines maintain the use of repletion dose of antioxidant micronutrients, and not the use of very high doses [[102](#page-9-0)].

### **Inflammation modulation using probiotics**

Dysbiosis is the disruption of the homeostasis of the microbiota. In ICU patients, intestinal dysbiosis is driven by numerous factors including antimicrobial treatment, the change in nutritional pattern, the splanchnic hypoperfusion and the stress of the acute illness [\[103\]](#page-9-37). Emergence of a pathobiome (reduced diversity and changes to a more pathogenic flora) is observed within hours following varied insults such as trauma, sepsis or extended burns [[104\]](#page-9-38). Alterations in intestinal homeostasis and gut microbiota in critical illness have been associated with increased inflammatory cytokine production [[105\]](#page-9-39), possibly due to the passage of microbial products in the circulation via the mesenteric lymph. The repletion of the gut with health-promoting probiotics (living microbes of human origin), potentially combined with prebiotics (synbiotics), has been hypothesized as a promising strategy to maintain the intestinal homeostasis. A recent meta-analysis including 33 studies examined the efficacy of probiotics or synbiotics in critically ill patients and concluded they could significantly reduce the incidence of ventilation associated pneumonia, and decrease the ICU length of stay and mortality as well [[106](#page-9-40)].

The impact of probiotics or symbiotics administration on the pro-inflammatory/anti-inflammatory balance in critically ill patients is unclear [[107](#page-9-41)]. Small studies showed interesting effects of synbiotics on different inflammation markers in various contexts such as unselected acute disease [[108\]](#page-9-42), severe acute pancreatitis [[109\]](#page-9-43) or hepatectomy [[110](#page-9-44)].

Importantly, the studies about microbiota-targeted therapies suffer from a huge heterogeneity in both patients and compounds tested. Therefore, drawing firm conclusions remains hazardous [\[111\]](#page-9-45). Moreover, concerns regarding the safety of probiotics exist, especially in the context of acute critical illness [\[112](#page-9-46)]. Due to their immunocompromised status, critically ill patients are at risk of bacteremia or fungemia secondary to probiotics administration [[113](#page-9-47)].

In the future, microbiome modulation may become a novel adjuvant strategy in inflammation modulation [[114\]](#page-9-48). A final aim could be to prevent or even treat ICUrelated sarcopenia and progression to chronic critical illness. There is promising research underway that should contribute to the understanding of how to use individualised microbiota-targeted treatment [[115\]](#page-9-49).

#### **Inflammation modulation using metabolic treatments**

The blockade of β-adrenergic stimulation with propranolol is used to blunt the catecholamine-mediated hypermetabolism. This effect has been confirmed in studies performed in burn patients during the acute phase as well as during rehabilitation [\[116,](#page-10-2) [117\]](#page-10-3). In severe burn children, propranolol given after 7 days following injury significantly decreased TNF-α and IL-1b. According to a recent metabolomic analysis, propranolol appears to mitigate some of the pathophysiologic effects of stress including lipidic metabolism, mitochondrial function and endoplasmic reticulum stress [\[118\]](#page-10-4). Evidence of benefits in non-burn ICU patients is currently lacking.

Oxandrolone, an oral synthetic testosterone analog, has been used for decades in acute and rehabilitating severe burn patients with beneficial effects on body composition and wound healing, while being safe for liver function [[119\]](#page-10-5). More recently, its effect on inflammatory response was investigated in a murine model of burn injury. Interestingly, oxandrolone treatment during the first days after injury led to a generalized reduction of pro-inflammatory mediators and an oxidative stress suppression in some organs  $[120]$ . Up to now, oxandrolone has never been studied in chronic critical illness or in ICU survivors. Unfortunately, oxandrolone is not available in some countries, but could then be substituted by nandrolone decanoate [\[121\]](#page-10-7).

Insulin at high dose is considered as an anabolic agent, by increasing muscle protein turnover. It is thought to act by increasing bioavailability of IGF1, and also by anti-inflammatory properties. Indeed, in severe burn patients, insulin treatment decreased expression of proinflammatory cytokines, increased anti-inflammatory cytokines, and modulated the synthesis of acute phase proteins [\[122](#page-10-8)]. Same findings were detected in non-burn critically ill patients with prolonged stay in ICU [[123\]](#page-10-9) In similar patients, intensive insulin therapy was also found to be associated with a decrease in cortisol blood level, independently of any effect on cortisol binding protein [[124\]](#page-10-10).However, the effective dose (around 0.0015 U/kg/ min) is greater than the one used to maintain euglycemia (glycemia<180 mg/dL) during acute critical illness, and exposes to a risk of hypoglycemia that can be mitigated by administration of exogenous glucose.

Metformin is an antihyperglycemic agent acting by decreasing hepatic glucose production while increasing muscle glucose uptake. Metformin has anti-inflammatory properties through inhibition of the NF-κB activation [\[125](#page-10-11)]. In patients with diabetes mellitus admitted in ICU for sepsis, preadmission metformin use may reduce mortality [\[126](#page-10-12)]. The underlying mechanism is unclear, but probably multifactorial in relation with the different properties of metformin: mitigation of inflammation, autophagy and mitochondrial functions [[127\]](#page-10-13). In

a model of mice suffering from sepsis, metformin treatment decreased the messenger ribonucleic acid levels of inflammatory factors and cytokines [[128\]](#page-10-14). Metformin has also been demonstrated to have anabolic properties in severe burn patients, probably as a consequence of glycemia control. Promising data were recently published in a mice model of thermal injury of moderate severity, showing a preservation of muscle mass and a proliferation of muscle satellite cells in treated animals [\[129](#page-10-15)]. The treatment is rarely associated with hypoglycemia and is now widely studied in non-diabetics [\[130\]](#page-10-16). However, its properties other than anti-hyperglycemic have never been studied in ICU survivors.

Melatonin, a pleiotropic hormone secreted by the pineal gland, plays roles in circadian rhythm, immune regulation and energy metabolism. In recent years, its administration as dietary supplement has gain popularity, especially for its protective role in sarcopenia related diseases. Indeed, among its protective effects on ICU-acquired weakness through reduction of proteolysis and improvement of mitochondrial dysfunction, melatonin has been shown, mainly in animal models, to have a direct anti-inflammatory effect  $[131]$  $[131]$ . Data on its use during acute illness or during rehabilitation aiming at enhancing muscle recovery are lacking, although this treatment seems to be safe at doses usually used for sleep promotion.

# **Conclusion**

Numerous strategies of inflammation attenuation, based on the physiologic effects of nutrients or pharmacological agents, are available in human clinical practice. Regretfully, there isn't always substantial proof from clinical research regarding the positive impacts that are directly brought about by inflammation modulation. Some arguments come from small studies performed in severe burn patients, but such results should be transposed to non-burn patients with caution. Given the metabolic effects of persisting inflammation and their long-term consequences, it makes sense to modulate it. However, the ideal timing for inflammation modulation is still undetermined, as well as the doses or the potential synergic effects of the different strategies. The ultimate goal is to individualize the modulation of inflammation, but this is made difficult by the imprecise or ambiguous clinical monitoring of inflammation that is available at the bedside. In the next future, further studies are needed to explore how the modulation of inflammation can improve the long-term outcomes after a critical illness.

#### **Abbreviations**





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#### **Author contributions**

AFR and RM wrote the manuscript. They both approved the final version of the manuscript.

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#### **References**

- <span id="page-7-0"></span>1. Mechanick JI, Brett EM. Endocrine and metabolic issues in the management of the chronically critically ill patient. Crit Care Clin. 2002;18:619–41. viii.
- <span id="page-7-1"></span>2. Knuth CM, Auger C, Chi L, et al. Thermal stress induces long-term remodeling of adipose tissue and is Associated with systemic dysfunction. Shock. 2021;56:744–54.
- <span id="page-7-2"></span>3. Pugin J. How tissue injury alarms the immune system and causes a systemic inflammatory response syndrome. Ann Intensiv Care. 2012;2:27.
- <span id="page-7-3"></span>4. Kavazis AN, Talbert EE, Smuder AJ, et al. Mechanical ventilation induces diaphragmatic mitochondrial dysfunction and increased oxidant production. Free Radic Biol Med. 2009;46:842–50.
- <span id="page-7-4"></span>5. Barnig C, Bezema T, Calder PC, et al. Activation of Resolution pathways to prevent and fight chronic inflammation: lessons from Asthma and Inflammatory Bowel Disease. Front Immunol. 2019;10:1699.
- <span id="page-7-5"></span>6. Hawkins RB, Raymond SL, Stortz JA, et al. Chronic critical illness and the persistent inflammation, immunosuppression, and catabolism syndrome. Front Immunol. 2018;9:1511.
- <span id="page-7-6"></span>7. Bagshaw SM, Stelfox HT, Iwashyna TJ, et al. Timing of onset of persistent critical illness: a multi-centre retrospective cohort study. Intensive Care Med. 2018;44:2134–44.
- <span id="page-7-7"></span>8. Moore FA, Phillips SM, McClain CJ, et al. Nutrition support for persistent inflammation, immunosuppression, and catabolism syndrome. Nutr Clin Pract. 2017;32:S121–7.
- <span id="page-7-8"></span>9. Committee IPG, Steering S, Advisory S. ISBI practice guidelines for burn Care. Burns. 2016;42:953–1021.
- <span id="page-7-9"></span>10. Jeschke MG, Gauglitz GG, Kulp GA, et al. Long-term persistance of the pathophysiologic response to severe burn injury. PLoS ONE. 2011;6:e21245.
- <span id="page-7-10"></span>11. Whittle J, Molinger J, MacLeod D, et al. Persistent hypermetabolism and longitudinal energy expenditure in critically ill patients with COVID-19. Crit Care. 2020;24:581.
- <span id="page-8-0"></span>12. Joris M, Minguet P, Colson C, et al. Cardiopulmonary Exercise Testing in critically ill coronavirus Disease 2019 survivors: evidence of a sustained Exercise Intolerance and Hypermetabolism. Crit Care Explor. 2021;3:e0491.
- <span id="page-8-1"></span>13. Joris M, Pincemail J, Colson C et al. Exercise limitation after critical versus mild COVID-19 infection: a metabolic perspective. J Clin Med 2022;11.
- <span id="page-8-2"></span>14. Griffith DM, Lewis S, Rossi AG, et al. Systemic inflammation after critical illness: relationship with physical recovery and exploration of potential mechanisms. Thorax. 2016;71:820–9.
- <span id="page-8-3"></span>15. Ogunbileje JO, Porter C, Herndon DN, et al. Hypermetabolism and hypercatabolism of skeletal muscle accompany mitochondrial stress following severe burn trauma. Am J Physiol Endocrinol Metabolism. 2016;311:E436–448.
- <span id="page-8-4"></span>16. Auger C, Samadi O, Jeschke MG. The biochemical alterations underlying post-burn hypermetabolism. Biochim Biophys Acta Mol Basis Dis. 2017;1863:2633–44.
- <span id="page-8-5"></span>17. Rousseau AF, Prescott HC, Brett SJ, et al. Long-term outcomes after critical illness: recent insights. Crit Care. 2021;25:108.
- <span id="page-8-6"></span>18. Docherty C, Page C, Wilson J et al. Association between inflammation and post-intensive care syndrome: a systematic review. Anaesthesia 2024.
- <span id="page-8-7"></span>19. Braga M, Wischmeyer PE, Drover J, et al. Clinical evidence for pharmaconutrition in major elective surgery. JPEN J Parenter Enter Nutr. 2013;37:S66–72.
- <span id="page-8-8"></span>20. Gianotti L, Nespoli L, Sandini M. Pharmaconutrition: which substrates? Eur J Surg Oncol 2022.
- <span id="page-8-9"></span>21. Eldaly AS, Avila FR, Torres R, et al. Modulation of burn Hypermetabolism in Preclinical models. Cureus. 2023;15:e33518.
- <span id="page-8-10"></span>22. Long CL, Schaffel N, Geiger JW, et al. Metabolic response to injury and illness: estimation of energy and protein needs from indirect calorimetry and nitrogen balance. JPEN J Parenter Enter Nutr. 1979;3:452–6.
- <span id="page-8-11"></span>23. Porter C, Tompkins RG, Finnerty CC, et al. The metabolic stress response to burn trauma: current understanding and therapies. Lancet. 2016;388:1417–26.
- <span id="page-8-12"></span>24. Jeschke MG, Chinkes DL, Finnerty CC, et al. Pathophysiologic response to severe burn injury. Ann Surg. 2008;248:387–401.
- <span id="page-8-13"></span>25. Bateman AP, McArdle F, Walsh TS. Time course of anemia during six months follow up following intensive care discharge and factors associated with impaired recovery of erythropoiesis. Crit Care Med. 2009;37:1906–12.
- <span id="page-8-14"></span>26. Vicic VK, Radman M, Kovacic V. Early initiation of enteral nutrition improves outcomes in burn disease. Asia Pac J Clin Nutr. 2013;22:543–7.
- <span id="page-8-15"></span>27. Gottschlich MM, Jenkins ME, Mayes T, et al. The 2002 Clinical Research Award. An evaluation of the safety of early vs delayed enteral support and effects on clinical, nutritional, and endocrine outcomes after severe burns. J Burn Care Rehabil. 2002;23:401–15.
- <span id="page-8-16"></span>28. Peck MD, Kessler M, Cairns BA, et al. Early Enteral Nutrition does not decrease Hypermetabolism Associated with burn Injury. J Trauma: Injury Infect Crit Care. 2004;57:1143–9.
- <span id="page-8-19"></span>29. Berger MM, Pantet O, Jacquelin-Ravel N, et al. Supplemental parenteral nutrition improves immunity with unchanged carbohydrate and protein metabolism in critically ill patients: the SPN2 randomized tracer study. Clin Nutr. 2019;38:2408–16.
- <span id="page-8-20"></span>30. Liu P, Wang Y, Yang G, et al. The role of short-chain fatty acids in intestinal barrier function, inflammation, oxidative stress, and colonic carcinogenesis. Pharmacol Res. 2021;165:105420.
- <span id="page-8-21"></span>31. Merker M, Felder M, Gueissaz L, et al. Association of baseline inflammation with effectiveness of nutritional support among patients with Diseaserelated malnutrition: a secondary analysis of a Randomized Clinical Trial. JAMA Netw Open. 2020;3:e200663.
- <span id="page-8-22"></span>32. Wischmeyer PE. Glutamine: mode of action in critical illness. Crit Care Med. 2007;35:S541–544.
- <span id="page-8-23"></span>33. Wischmeyer PE. Glutamine in burn Injury. Nutr Clin Pract. 2019;34:681–7.
- <span id="page-8-24"></span>34. Wang ZE, Zheng JJ, Bin Feng J, et al. Glutamine relieves the hypermetabolic response and reduces organ damage in severe burn patients: a multicenter, randomized controlled clinical trial. Burns. 2022;48:1606–17.
- <span id="page-8-25"></span>35. Heyland DK, Wibbenmeyer L, Pollack JA, et al. A Randomized Trial of Enteral glutamine for treatment of burn injuries. N Engl J Med. 2022;387:1001–10.
- <span id="page-8-26"></span>36. Berger MM, Binz PA, Roux C, et al. Exudative glutamine losses contribute to high needs after burn injury. JPEN J Parenter Enter Nutr. 2022;46:782–8.
- <span id="page-8-18"></span>37. Singer P, Blaser AR, Berger MM, et al. ESPEN practical and partially revised guideline: clinical nutrition in the intensive care unit. Clin Nutr. 2023;42:1671–89.
- <span id="page-8-17"></span>38. Singer P, Blaser AR, Berger MM, et al. ESPEN guideline on clinical nutrition in the intensive care unit. Clin Nutr. 2019;38:48–79.
- <span id="page-8-27"></span>39. Heyland D, Muscedere J, Wischmeyer PE, et al. A randomized trial of glutamine and antioxidants in critically ill patients. N Engl J Med. 2013;368:1489–97.
- <span id="page-8-28"></span>40. Heyland DK, Elke G, Cook D, et al. Glutamine and antioxidants in the critically ill patient: a post hoc analysis of a large-scale randomized trial. JPEN J Parenter Enter Nutr. 2015;39:401–9.
- <span id="page-8-29"></span>41. van Zanten AR, Dhaliwal R, Garrel D, et al. Enteral glutamine supplementation in critically ill patients: a systematic review and meta-analysis. Crit Care. 2015;19:294.
- <span id="page-8-30"></span>42. Smedberg M, Wernerman J. Is the glutamine story over? Crit Care. 2016;20:361.
- <span id="page-8-31"></span>43. MacLeod AS, Mansbridge JN. The Innate Immune System in Acute and Chronic wounds. Adv Wound Care (New Rochelle). 2016;5:65–78.
- <span id="page-8-32"></span>44. Veglia F, Perego M, Gabrilovich D. Myeloid-derived suppressor cells coming of age. Nat Immunol. 2018;19:108–19.
- <span id="page-8-33"></span>45. Zhu X, Pribis JP, Rodriguez PC, et al. The central role of arginine catabolism in T-cell dysfunction and increased susceptibility to infection after physical injury. Ann Surg. 2014;259:171–8.
- 46. McCarthy MS, Martindale RG. Immunonutrition in critical illness: what is the role? Nutr Clin Pract. 2018;33:348–58.
- <span id="page-8-34"></span>47. Drover JW, Dhaliwal R, Weitzel L, et al. Perioperative use of argininesupplemented diets: a systematic review of the evidence. J Am Coll Surg. 2011;212:385–99. 399 e381.
- <span id="page-8-35"></span>48. Rosenthal MD, Carrott PW, Patel J, et al. Parenteral or Enteral Arginine Supplementation Safety and Efficacy. J Nutr. 2016;146:S2594–600.
- <span id="page-8-36"></span>49. Luiking YC, Poeze M, Deutz NE. Arginine infusion in patients with septic shock increases nitric oxide production without haemodynamic instability. Clin Sci (Lond). 2015;128:57–67.
- <span id="page-8-37"></span>50. Luiking YC, Poeze M, Deutz NE. A randomized-controlled trial of arginine infusion in severe sepsis on microcirculation and metabolism. Clin Nutr. 2020;39:1764–73.
- <span id="page-8-38"></span>51. Agarwal U, Didelija IC, Yuan Y, et al. Supplemental Citrulline is more efficient than arginine in increasing systemic arginine availability in mice. J Nutr. 2017;147:596–602.
- <span id="page-8-39"></span>52. Tadie JM, Locher C, Maamar A, et al. Enteral citrulline supplementation versus placebo on SOFA score on day 7 in mechanically ventilated critically ill patients: the IMMUNOCITRE randomized clinical trial. Crit Care. 2023;27:381.
- <span id="page-8-40"></span>Bonafe L, Berger MM, Que YA, et al. Carnitine deficiency in chronic critical illness. Curr Opin Clin Nutr Metab Care. 2014;17:200–9.
- <span id="page-8-41"></span>54. Berger MM, Broman M, Forni L, et al. Nutrients and micronutrients at risk during renal replacement therapy: a scoping review. Curr Opin Crit Care. 2021;27:367–77.
- <span id="page-8-42"></span>55. Rousseau AF, Schmitz S, Cavalier E et al. Altered serum acylcarnitines Profile after a prolonged stay in Intensive Care. Nutrients 2022;14.
- 56. Rousseau AF, Dongier A, Colson C, et al. Serum acylcarnitines Profile in critically ill survivors according to illness severity and ICU length of Stay: an observational study. Nutrients. 2023;15:2392.
- <span id="page-8-43"></span>57. Rousseau AF, Ngongan A, Colson C et al. Mid-term evolution of the serum acylcarnitine Profile in critically ill survivors: a metabolic insight into Survivorship. Nutrients 2023;15.
- <span id="page-8-44"></span>58. Yahyapoor F, Sedaghat A, Feizi A, et al. The effects of l-Carnitine supplementation on inflammatory markers, clinical status, and 28 days mortality in critically ill patients: a double-blind, randomized, placebo-controlled trial. Clin Nutr ESPEN. 2022;49:61–7.
- <span id="page-8-45"></span>59. Keshani M, Alikiaii B, Babaei Z, et al. The effects of L-carnitine supplementation on inflammation, oxidative stress, and clinical outcomes in critically ill patients with sepsis: a randomized, double-blind, controlled trial. Nutr J. 2024;23:31.
- <span id="page-8-46"></span>60. Haghighatdoost F, Jabbari M, Hariri M. The effect of L-carnitine on inflammatory mediators: a systematic review and meta-analysis of randomized clinical trials. Eur J Clin Pharmacol. 2019;75:1037–46.
- <span id="page-8-47"></span>61. Fielding R, Riede L, Lugo JP et al. l-Carnitine Supplementation in Recovery after Exercise. *Nutrients* 2018;10.
- <span id="page-8-48"></span>62. Calder PC. Omega-3 polyunsaturated fatty acids and inflammatory processes: nutrition or pharmacology? Br J Clin Pharmacol. 2013;75:645–62.
- <span id="page-8-49"></span>63. Manzanares W, Langlois PL, Dhaliwal R, et al. Intravenous fish oil lipid emulsions in critically ill patients: an updated systematic review and meta-analysis. Crit Care. 2015;19:167.
- <span id="page-8-50"></span>64. Singer P, Calder PC. The role of omega-3 polyunsaturated fatty acids in the intensive care unit. Curr Opin Clin Nutr Metab Care. 2023;26:129–37.
- <span id="page-9-1"></span>66. Huang Z, Zheng J, Huang W, et al. The effects and safety of omega-3 fatty for acute lung injury: a systematic review and meta-analysis. World J Surg Oncol. 2020;18:235.
- <span id="page-9-2"></span>67. Kristine Koekkoek W, Panteleon V, van Zanten AR. Current evidence on omega-3 fatty acids in enteral nutrition in the critically ill: a systematic review and meta-analysis. Nutrition. 2019;59:56–68.
- <span id="page-9-3"></span>68. Wang H, Su S, Wang C, et al. Effects of fish oil-containing nutrition supplementation in adult sepsis patients: a systematic review and meta-analysis. Burns Trauma. 2022;10:tkac012.
- <span id="page-9-4"></span>69. Wang C, Han D, Feng X, et al. Omega-3 fatty acid supplementation is associated with favorable outcomes in patients with sepsis: an updated metaanalysis. J Int Med Res. 2020;48:300060520953684.
- <span id="page-9-5"></span>70. Serhan CN, Chiang N. Resolvins and cysteinyl-containing pro-resolving mediators activate resolution of infectious inflammation and tissue regeneration. Prostaglandins Other Lipid Mediat. 2023;166:106718.
- <span id="page-9-6"></span>71. Di Girolamo FG, Situlin R, Mazzucco S, et al. Omega-3 fatty acids and protein metabolism: enhancement of anabolic interventions for Sarcopenia. Curr Opin Clin Nutr Metab Care. 2014;17:145–50.
- <span id="page-9-7"></span>72. Bird JK, Troesch B, Warnke I, et al. The effect of long chain omega-3 polyunsaturated fatty acids on muscle mass and function in Sarcopenia: a scoping systematic review and meta-analysis. Clin Nutr ESPEN. 2021;46:73–86.
- <span id="page-9-8"></span>73. Abribat T, Nedelec B, Jobin N, et al. Decreased serum insulin-like growth factor-I in burn patients: relationship with serum insulin-like growth factor binding protein-3 proteolysis and the influence of lipid composition in nutritional support. Crit Care Med. 2000;28:2366–72.
- <span id="page-9-9"></span>74. Serhan CN, Back M, Chiurchiu V, et al. Expert consensus report on lipid mediators: role in resolution of inflammation and muscle preservation. FASEB J. 2024;38:e23699.
- <span id="page-9-10"></span>75. Kottler ML. Is vitamin D a key factor in muscle health? Endocrinology. 2013;154:3963–4.
- <span id="page-9-11"></span>76. Ahuja A, Agrawal S, Acharya S, et al. A Comprehensive Review of the Immunomodulatory effects of vitamin D in Sepsis. Cureus. 2024;16:e53678.
- <span id="page-9-12"></span>77. Ryan ZC, Craig TA, Folmes CD, et al. 1alpha,25-Dihydroxyvitamin D3 regulates mitochondrial Oxygen Consumption and Dynamics in human skeletal muscle cells. J Biol Chem. 2016;291:1514–28.
- <span id="page-9-13"></span>78. Cavalier E, Makris K, Heijboer AC et al. Vitamin D: Analytical advances, clinical impact, and Ongoing debates on Health perspectives. Clin Chem 2024.
- <span id="page-9-14"></span>79. Mansournia MA, Ostadmohammadi V, Doosti-Irani A, et al. The effects of vitamin D supplementation on biomarkers of inflammation and oxidative stress in Diabetic patients: a systematic review and Meta-analysis of Randomized controlled trials. Horm Metab Res. 2018;50:429–40.
- <span id="page-9-15"></span>80. Gwenzi T, Zhu A, Schrotz-King P, et al. Effects of vitamin D supplementation on inflammatory response in patients with cancer and precancerous lesions: systematic review and meta-analysis of randomized trials. Clin Nutr. 2023;42:1142–50.
- <span id="page-9-16"></span>81. Quraishi SA, De Pascale G, Needleman JS, et al. Effect of Cholecalciferol supplementation on vitamin D status and Cathelicidin Levels in Sepsis: a Randomized, Placebo-Controlled Trial. Crit Care Med. 2015;43:1928–37.
- <span id="page-9-17"></span>82. Berger MM, Amrein K, Barazzoni R et al. The science of micronutrients in clinical practice - Report on the ESPEN symposium. *Clin Nutr* 2024;43:268–283.
- <span id="page-9-18"></span>83. Klein GL, Chen TC, Holick MF, et al. Synthesis of vitamin D in skin after burns. Lancet. 2004;363:291–2.
- <span id="page-9-19"></span>84. Holick MF, Binkley NC, Bischoff-Ferrari HA, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2011;96:1911–30.
- <span id="page-9-20"></span>85. Menger J, Lee ZY, Notz Q, et al. Administration of vitamin D and its metabolites in critically ill adult patients: an updated systematic review with metaanalysis of randomized controlled trials. Crit Care. 2022;26:268.
- <span id="page-9-21"></span>86. Amrein K, Schnedl C, Holl A, et al. Effect of high-dose vitamin D3 on hospital length of stay in critically ill patients with vitamin D deficiency: the VITdAL-ICU randomized clinical trial. JAMA. 2014;312:1520–30.
- <span id="page-9-22"></span>87. Bouillon R, Manousaki D, Rosen C, et al. The health effects of vitamin D supplementation: evidence from human studies. Nat Rev Endocrinol. 2022;18:96–110.
- <span id="page-9-23"></span>88. Griffin G, Hewison M, Hopkin J, et al. Perspective: vitamin D supplementation prevents rickets and acute respiratory infections when given as daily maintenance but not as intermittent bolus: implications for COVID-19. Clin Med (Lond). 2021;21:e144–9.
- <span id="page-9-24"></span>89. Manzanares W, Dhaliwal R, Jiang X, et al. Antioxidant micronutrients in the critically ill: a systematic review and meta-analysis. Crit Care. 2012;16:R66.
- <span id="page-9-25"></span>90. Dresen E, Pimiento JM, Patel JJ, et al. Overview of oxidative stress and the role of micronutrients in critical illness. JPEN J Parenter Enter Nutr. 2023;47(Suppl 1):S38–49.
- <span id="page-9-26"></span>91. Koekkoek WA, van Zanten AR. Antioxidant vitamins and Trace Elements in critical illness. Nutr Clin Pract. 2016;31:457–74.
- <span id="page-9-27"></span>92. Gudivada KK, Kumar A, Sriram K, et al. Antioxidant micronutrient supplements for adult critically ill patients: a bayesian multiple treatment comparisons meta-analysis. Clin Nutr ESPEN. 2022;47:78–88.
- <span id="page-9-28"></span>93. Berger MM, Talwar D, Shenkin A. Pitfalls in the interpretation of blood tests used to assess and monitor micronutrient nutrition status. Nutr Clin Pract. 2023;38:56–69.
- <span id="page-9-29"></span>94. Tanaka H, Matsuda T, Miyagantani Y, et al. Reduction of resuscitation fluid volumes in severely burned patients using ascorbic acid administration: a randomized, prospective study. Arch Surg. 2000;135:326–31.
- <span id="page-9-30"></span>95. Tanaka H. Regarding the efficacy of postburn vitamin C infusions on edema formation. J Burn Care Rehabil. 1999;20:437–8.
- <span id="page-9-31"></span>96. Patel JJ, Ortiz-Reyes A, Dhaliwal R, et al. IV vitamin C in critically ill patients: a systematic review and Meta-analysis. Crit Care Med. 2022;50:e304–12.
- <span id="page-9-32"></span>97. Lee ZY, Ortiz-Reyes L, Lew CCH, et al. Intravenous vitamin C monotherapy in critically ill patients: a systematic review and meta-analysis of randomized controlled trials with trial sequential analysis. Ann Intensiv Care. 2023;13:14.
- <span id="page-9-33"></span>98. Berger MM, Soguel L, Shenkin A, et al. Influence of early antioxidant supplements on clinical evolution and organ function in critically ill cardiac surgery, major trauma, and subarachnoid hemorrhage patients. Crit Care. 2008;12:R101.
- <span id="page-9-34"></span>99. Fowler AA 3rd, Syed AA, Knowlson S, et al. Phase I safety trial of intravenous ascorbic acid in patients with severe sepsis. J Translational Med. 2014;12:32.
- <span id="page-9-35"></span>100. Fujii T, Lankadeva YR, Bellomo R. Update on vitamin C administration in critical illness. Curr Opin Crit Care. 2022;28:374–80.
- <span id="page-9-36"></span>101. Mahmoodpoor A, Faramarzi E, Reyhanifard A, et al. The effects of selenium supplementation on inflammatory markers in critically ill patients. SN Appl Sci. 2022;4:326.
- <span id="page-9-0"></span>102. Berger MM, Shenkin A, Schweinlin A, et al. ESPEN micronutrient guideline. Clin Nutr. 2022;41:1357–424.
- <span id="page-9-37"></span>103. Wozniak H, Beckmann TS, Frohlich L, et al. The central and biodynamic role of gut microbiota in critically ill patients. Crit Care. 2022;26:250.
- <span id="page-9-38"></span>104. McDonald D, Ackermann G, Khailova L et al. Extreme Dysbiosis Microbiome Crit Illn mSphere 2016;1.
- <span id="page-9-39"></span>105. Wischmeyer PE, McDonald D, Knight R. Role of the microbiome, probiotics, and 'dysbiosis therapy' in critical illness. Curr Opin Crit Care. 2016;22:347–53.
- <span id="page-9-40"></span>106. Lou J, Cui S, Huang N, et al. Efficacy of probiotics or synbiotics in critically ill patients: a systematic review and meta-analysis. Clin Nutr ESPEN. 2024;59:48–62.
- <span id="page-9-41"></span>107. Giron M, Thomas M, Dardevet D, et al. Gut microbes and muscle function: can probiotics make our muscles stronger? J cachexia Sarcopenia Muscle. 2022;13:1460–76.
- <span id="page-9-42"></span>108. Seifi N, Sedaghat A, Nematy M, et al. Effects of synbiotic supplementation on the serum endotoxin level, inflammatory status, and clinical outcomes of adult patients with critical illness: a randomized controlled trial. Nutr Clin Pract. 2022;37:451–8.
- <span id="page-9-43"></span>109. Rohith G, Sureshkumar S, Anandhi A, et al. Effect of Synbiotics in reducing the systemic inflammatory response and septic complications in moderately severe and severe Acute Pancreatitis: a prospective parallel-arm double-blind Randomized Trial. Dig Dis Sci. 2023;68:969–77.
- <span id="page-9-44"></span>110. Sugawara G, Nagino M, Nishio H, et al. Perioperative synbiotic treatment to prevent postoperative infectious complications in biliary cancer surgery: a randomized controlled trial. Ann Surg. 2006;244:706–14.
- <span id="page-9-45"></span>111. Biemond JJ, McDonald B, Haak BW. Leveraging the microbiome in the treatment of sepsis: potential pitfalls and new perspectives. Curr Opin Crit Care. 2023;29:123–9.
- <span id="page-9-46"></span>112. Cohen PA. Probiotic Safety-No guarantees. JAMA Intern Med. 2018;178:1577–8.
- <span id="page-9-47"></span>113. Didari T, Solki S, Mozaffari S, et al. A systematic review of the safety of probiotics. Expert Opin Drug Saf. 2014;13:227–39.
- <span id="page-9-48"></span>114. Shimizu K, Ojima M, Ogura H. Gut microbiota and Probiotics/Synbiotics for Modulation of Immunity in critically ill patients. Nutrients 2021;13.
- <span id="page-9-49"></span>115. Haak BW, Prescott HC, Wiersinga WJ. Therapeutic potential of the gut microbiota in the Prevention and Treatment of Sepsis. Front Immunol. 2018;9:2042.
- <span id="page-10-2"></span>116. Herndon DN, Rodriguez NA, Diaz EC, et al. Long-term Propranolol Use in severely burned Pediatric patients: a randomized controlled study. Ann Surg. 2012;256:402–11.
- <span id="page-10-3"></span>117. Herndon DN, Hart DW, Wolf SE, et al. Reversal of catabolism by beta-blockade after severe burns. N Engl J Med. 2001;345:1223–9.
- <span id="page-10-4"></span>118. Rehou S, de Brito Monteiro L, Auger C et al. Propranolol normalizes metabolomic signatures thereby improving outcomes after burn. Ann Surg 2023.
- <span id="page-10-5"></span>119. Ring J, Heinelt M, Sharma S, et al. Oxandrolone in the treatment of burn injuries: a systematic review and Meta-analysis. J Burn Care Res. 2020;41:190–9.
- <span id="page-10-6"></span>120. Ahmad A, Herndon DN, Szabo C. Oxandrolone protects against the development of multiorgan failure, modulates the systemic inflammatory response and promotes wound healing during burn injury. Burns. 2019;45:671–81.
- <span id="page-10-7"></span>121. Ali YH, Ali T. Nandrolone decanoate safely combats catabolism in burned patients: a new potential indication after recall. Burns. 2022;48:59–68.
- <span id="page-10-8"></span>122. Jeschke MG, Boehning DF, Finnerty CC, et al. Effect of insulin on the inflammatory and acute phase response after burn injury. Crit Care Med. 2007;35:S519–523.
- <span id="page-10-9"></span>123. Hansen TK, Thiel S, Wouters PJ, et al. Intensive insulin therapy exerts antiinflammatory effects in critically ill patients and counteracts the adverse effect of low mannose-binding lectin levels. J Clin Endocrinol Metab. 2003;88:1082–8.
- <span id="page-10-10"></span>124. Vanhorebeek I, Peeters RP, Vander Perre S, et al. Cortisol response to critical illness: effect of intensive insulin therapy. J Clin Endocrinol Metab. 2006;91:3803–13.
- <span id="page-10-11"></span>125. Saisho Y. Metformin and inflammation: its potential beyond glucose-lowering Effect. Endocr Metab Immune Disord Drug Targets. 2015;15:196–205.
- <span id="page-10-12"></span>126. Li Y, Zhao H, Guo Y, et al. Association of Preadmission Metformin Use and Prognosis in patients with Sepsis and diabetes Mellitus: a systematic review and Meta-analysis. Front Endocrinol (Lausanne). 2021;12:811776.
- <span id="page-10-13"></span>127. Bharath LP, Nikolajczyk BS. The intersection of metformin and inflammation. Am J Physiol Cell Physiol. 2021;320:C873–9.
- <span id="page-10-14"></span>128. Song H, Zhang X, Zhai R, et al. Metformin attenuated sepsis-associated liver injury and inflammatory response in aged mice. Bioengineered. 2022;13:4598–609.
- <span id="page-10-15"></span>129. Yousuf Y, Datu A, Barnes B, et al. Metformin alleviates muscle wasting postthermal injury by increasing Pax7-positive muscle progenitor cells. Stem Cell Res Ther. 2020;11:18.
- <span id="page-10-16"></span>130. Naseri A, Sanaie S, Hamzehzadeh S, et al. Metformin: new applications for an old drug. J Basic Clin Physiol Pharmacol. 2023;34:151–60.
- <span id="page-10-17"></span>131. Liu Y, Wang D, Li T, et al. Melatonin: a potential adjuvant therapy for septic myopathy. Biomed Pharmacother. 2023;158:114209.
- <span id="page-10-0"></span>132. Rousseau AF, Losser MR, Ichai C, et al. ESPEN endorsed recommendations: Nutritional therapy in major burns. Clin Nutr. 2013;32:497–502.
- <span id="page-10-1"></span>133. Giustina A, Bilezikian JP, Adler RA et al. Consensus Statement on vitamin D Status Assessment and Supplementation: Whys, Whens, and Hows. Endocr Rev 2024.

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